

**THE ROLE OF GLUTATHIONE TRANSFERASES IN
RENAL CELL CARCINOMA****ULOGA GLUTATION TRANSFERAZA U KARCINOMU
BUBREŽNOG PARENHIMA**Vesna Ćorić¹, Marija Plješa-Ercegovac¹, Zoran Džamić²¹ University of Belgrade, Faculty of Medicine, Institute of Biochemistry, Belgrade, Serbia² University of Belgrade, Faculty of Medicine, Clinical Center of Serbia, Urological Clinic, Serbia**Correspondence:** coricmvesna@gmail.com**ABSTRACT**

Mounting evidence suggest that members of the subfamily of cytosolic glutathione S-transferases (GSTs) possess roles far beyond the classical glutathione-dependent enzymatic conjugation of electrophilic metabolites and xenobiotics. Namely, monomeric forms of certain GSTs are capable of forming protein: protein interactions with protein kinases and regulate cell apoptotic pathways. Due to this dual functionality of cytosolic GSTs, they might be implicated in both the development and the progression of renal cell carcinoma (RCC).

Prominent genetic heterogeneity, resulting from the gene deletions, as well as from SNPs in the coding and non-coding regions of GST genes, might affect GST isoenzyme profiles in renal parenchyma and therefore serve as a valuable indicator for predicting the risk of cancer development. Namely, GSTs are involved in the biotransformation of several compounds recognized as risk factors for RCC. The most potent carcinogen of polycyclic aromatic hydrocarbon diol epoxides, present in cigarette smoke, is of benzo(a)pyrene (BPDE), detoxified by GSTs. So far, the relationship between GST genotype and BPDE-DNA adduct formation, in determining the risk for RCC, has not been evaluated in patients with RCC.

Although the association between certain individual and combined GST genotypes and RCC risk has been debated in the literature, the data on the prognostic value of GST polymorphism in patients with RCC are scarce, probably due to the fact that the molecular mechanism supporting the role of GSTs in RCC progression has not been clarified as yet.

Keywords:Glutathione S-transferase,
polymorphism,
renal cell carcinoma

SAŽETAK

Veliki broj dokaza govori u prilog tome da pojedini pripadnici citosolnih glutathion S-transferaza (GST) poseduju i uloge nezavisne od njihove klasične uloge u konjugaciji elektrofilnih metabolita i ksenobiotika sa glutathionom. Naime, monomerni oblici pojedinih GST sposobni su da formiraju protein: proteinske interakcije sa izvesnim protein-kinazama i time regulišu puteve proliferacije i preživljavanja u ćeliji. Smatra se da zbog ove svoje dvostruke uloge citosolne GST mogu da utiču kako na nastanak, tako i na progresiju karcinoma bubrežnog parenhima (KBP).

Značajna genetska heterogenost, nastala ili kao rezultat delecije gena ili usled prisustva polimorfizma jednog nukleotida, kako u kodirajućim, tako i nekodirajućim sekvencama GST gena, može da utiče na GST izoenzimski profil u bubrežnom parenhimu i posluži kao dragoceni pokazatelj za procenu rizika za nastanak karcinoma. Glutathion transferaze su uključene u reakcije biotransformacije nekoliko jedinjenja priznatih kao faktori rizika za KBP. Benzo(a)piren diol-epoksid (BPDE) pripada grupi diol-epoksida iz grupe policikličnih aromatičnih ugljovodonika i, kao jedan od najopasnijih kancerogena prisutnih u duvanskom dimu, supstrat je za pojedine GST. Odnos između GST genotipa i nivoa BPDE-DNK adukta do sada nije analiziran u svetlu procene rizika za nastanak KBP.

Ključne reči:

glutathion S-transferaza, polimorfizam, karcinom bubrežnog parenhima

Iako je veza između određenih pojedinačnih i kombinovanih GST genotipova i rizika za nastanak KBP bila predmet analiza velikog broja radova, nema puno podataka koji govore u prilog prognostičkom značaju GST polimorfizma kod pacijenata sa KBP, verovatno zbog činjenice da molekularni mehanizam, koji je u osnovi uloge GST u progresiji KBP, nije još uvek razjašnjen.

RENAL CELL CARCINOMA (RCC)

Renal cell carcinoma (RCC) is the predominant form of kidney malignancy, comprising a group of heterogeneous renal tumors (1,2), with the clear cell RCC (ccRCC) being the most frequent subtype of sporadic RCC in adults (70-85%) (3,4).

In 2013, kidneys were recognized as the seventh most common site for tumor development (5). Renal cell carcinoma is the predominant form of kidney malignancy, whereas urothelial carcinoma, arising in the renal pelvis, accounts for less than 10% of histologically confirmed kidney carcinomas. (2). In 2012, the global incidence rate reported for RCC was 6.0/100.000 for men and 3.0/100.000 for women (5). Similarly, in 2013 the incidence in Serbia was reported as 6.1 (men) and 3.0 (women) per 100.000 people (6).

Most RCC are asymptomatic. It seems that the use of high-resolution cross-sectional imaging modalities over the last few decades has led to the increase in incidental detection of renal masses, often characterized as small, and low-graded (7). Nowadays, between 48-66% of such RCCs are detected incidentally (8). Still, many renal masses remain asymptomatic until the late stages of the disease. Despite advances in diagnostic methods, about 20-30% of patients are diagnosed with metastatic disease and 20% of patients undergoing nephrectomy will eventually develop metastatic RCC during the follow up period (9,10).

Cigarette smoking, obesity and hypertension are the most well established risk factors for sporadic RCC (2,10-13). Cigarette smoke is a rich source of free radicals, which are believed to be responsible for initiation of many tumors by inducing DNA damage that accumulates in

cells. In addition to free radicals, more than 60 carcinogens have been found in cigarette smoke. Among these, sufficient evidence of carcinogenicity was found for polycyclic aromatic hydrocarbons (PAHs), such as benzo(a)pyrene and aromatic amines, such as 4-amino biphenyl (14). Particular interest has been given to the most abundant, benzo(a)pyrene (B(a)P) and its carcinogenic metabolites, stereoisomers of 7,8-dihydroxy-9,10-oxo-7,8,9,10-tetrahydrobenzo(a)pyrene (BPDE) (15). The (+)-antiisomer [(+)-anti-BPDE] seems to be the most potent carcinogen of all PAH diol epoxides (16). Namely, BPDE is known as 'the bullet of the smoking gun', leaving its fingerprints in the blood of smokers, in the form of adducts with either serum albumins or DNA (15,17).

In recent years, the genetic origin of RCC became a focus of research, since not all individuals exposed to recognized RCC risk factors develop renal cell carcinoma. In general, an effort has been made towards identifying the common genetic variations, known as "quantitative trait loci", that could contribute a small, but significant risk not only for the development, but for the progression of complex disorder such as cancer (18).

GLUTATHIONE S- TRANSFERASES (GSTs)

A growing number of genes encoding enzymes involved in biotransformation and cellular defense has been identified, leading to increased knowledge of allelic variants of genes that may result in a differential susceptibility to environmental and oxidative stress (19,20). Glutathione transferases (EC 2.5.1.18), also referred to as glutathione

S-transferases or GSTs, are members of a multi-gene family. There are three major families of GST proteins, with cytosolic GSTs constituting the largest family (21). Seven classes of cytosolic GSTs have been identified in mammals (22), comprising a set of cellular proteins (GSTome) with various catalytic and non-catalytic functions (23,24). Namely, primary metabolic role of GST is to detoxify reactive electrophiles, such as potent xenobiotic, carcinogens and even therapeutic drugs (20), by catalyzing reaction of conjugation with glutathione. Glutathione conjugates are, thereafter, exported from the cell and subjected to metabolism of mercapturic acid, followed by the excretion in the urine (25) or bile (26). Thereby, GSTs reduce the likelihood of deleterious interactions of reactive compounds with important cellular macromolecules, such as proteins and nucleic acids (27).

However, not all reactions catalyzed by GST enzymes result in detoxification. Namely, in certain instances some GSTs are associated with bio-activation of electrophilic compounds (28,29) where the glutathione conjugate is more reactive than the parent compound. A growing number of evidence supports the aforementioned phenomenon, where mutagens, carcinogens and even some prodrugs are metabolically activated by conjugation with GSH (28,30). There are evidence that this is particularly true for the kidney (31,32).

Being a multifunctional group of enzymes, GSTs are involved in, intracellular binding and transport of hydrophobic compounds (33), and catalysis of key steps in the synthesis of leukotrienes, prostaglandins (34), steroid hormones (35), as well as the degradation of tyrosine (21). Moreover, some GST isoenzymes exhibit selenium independent glutathione peroxidase activity and along with other antioxidant enzymes provide a certain shield against a range of harmful electrophiles, produced during redox imbalance (36). Phospholipid, fatty acid and cholesterol hydroperoxides seem to be substrates for several GSTs, especially for the members of class *alpha* enzymes (37).

In addition to their role in the biotransformation reactions, there are evidence which clearly indicate the involvement of GST in the cellular survival, proliferation and apoptosis as well, by the means of protein: protein interactions with the signaling molecules, such as mitogen activating protein kinases (MAPK) (19,22,38,39). The first example of GST-mediated kinase regulation was the discovery of the GSTP1:JNK1 complexes (40). Namely, it seems that under physiological conditions, a portion of GSTP1 is bound to c-Jun NH2-terminal kinase (JNK1), regulating the level of JNK1 activity. However, in case of increased reactive oxygen species content, the GSTP1:JNK1 complex dissociates which in turn leads to the association of GSTP1 into oligomers. Now activated, JNK1 induces a chain of events, starting from the phosphorylation of its substrate, the transcription factor c-Jun, and resulting in apoptosis (19,40).

Another example of protein: protein interaction, similar to those of GSTP1, is a complex between mitogen

activated kinase (MAPK) ASK1 and GSTM1-1, found to be important for the maintenance of the normal level of p38 phosphorylation (41). Namely, ASK1 is MAPK kinase that activates JNK1 and p38 pathways, leading to cytokine and stressed-induced apoptosis (42). Environmental stress causes the disruption of the complex of GSTM1:ASK1, which accumulates GSTM1 into oligomers, while ASK1 is being activated (43). This dissociation results in a subsequent activation of JNK1 and p38-dependent signal pathways, ultimately leading to stress-induced apoptosis. In particular, *Cho et al., 2001* (41) showed both *in vitro* and *in vivo*, that mouse glutathione S-transferase Mu 1-1 (mGSTM1-1) physically interacts with ASK1 and, in doing so, functions as a negative regulator of ASK1 inside cells, repressing ASK1-mediated signals.

Genetic variations in human GSTs

Deletions and single-nucleotide polymorphisms (SNP) occur in genes encoding for members of the glutathione S-transferase superfamily (GSTs; EC 2.5.1.18), resulting in complete lack or alteration in enzyme activity (44).

Both *GSTM1* and *GSTT1* genes exhibit homozygous deletion polymorphisms, commonly referred to as *the null genotype*. The general lack of enzymes in such individuals has been recognized as potentially important modifier of individual risk for environmentally induced cancers (44). In case of *GSTM1-null* genotype, the underlying mechanism conferring an increased risk of cancer would be that such individuals are more susceptible to chemical-induced carcinogenesis, due to the diminished activity of xenobiotic-metabolizing defense system (45). On the other hand, it seems that when it comes to gene-environment interactions, *GSTT1* deficiency may be either deleterious or beneficial depending upon substrate exposure. Namely, members of the GST *theta* class are involved in bio-activation of certain compounds, producing even more toxic reactive intermediates, as a result of GSH conjugation (46).

Contrary to other GSTs, several SNPs have been identified in 5' non-coding promoter region of *GSTA1* gene, among them *GSTA1**C69T (rs3957356), reducing the levels of *GSTA1* enzyme in carriers of the variant *GSTA1**B in liver (Coles and Kadlubar, 2005). This *GSTA1* polymorphism is represented by three, apparently linked, SNPs: T-567G, C-69T and G-52A located within in the proximal promoter (47). It has been suggested that this genetic variation of *GSTA1* can change an individual's susceptibility to carcinogens and toxins, as well as affect the efficacy of some drugs (48).

GSTP1 SNP (rs1659) is one of the most extensively studied *GST* polymorphisms. This SNP encodes *the Ile105Val* substitution, which influences *Ile105* and *Val105* variants' catalytic efficacy (49,50) and has been investigated not only in terms of cancer susceptibility, but also in relation to drug resistance (22,51,52). It has been shown that *Ile105Val* substitution contributes to the architecture

of the hydrophobic substrate binding GST site and different substrate specificity (53). For instance, GSTP1 variants exhibit significantly different rates of conjugating activity towards (+)-anti-BPDE, with higher turnover for isoform GSTP1*Val105 than for isoform GSTP1*Ile105, due to the more favorable substrate-binding setting (50).

GENETIC POLYMORPHISM OF GLUTATHIONE TRANSFERASES IN PATIENTS WITH RENAL CELL CARCINOMA

A growing body of evidence suggests that cytosolic GSTs might be implicated not solely in the development, but also in the progression of RCC (54-57). The GSTs are involved in the biotransformation of several compounds recognized as risk factors for RCC (44). The main site for the initial glutathione conjugation of toxic compounds is generally assumed to be the liver, followed by a mandatory transfer of conjugates to the kidney (58). However, the initial bio-activation step of some nephrocarcinogens can take place in the kidney itself (59). The potential genotoxicity of carcinogens depends on the biotransformation capacity of renal tissue. Prominent genetic heterogeneity, resulting from the gene deletions, as well as from SNPs in the coding and non-coding regions of GST genes, might affect GST isoenzyme profile in renal parenchyma and, therefore, serve as a valuable indicator for predicting the risk of cancer development (45).

The deletion of *GSTM1* gene is one of the most investigated GST polymorphisms, since it has been suggested that a common variation within the *GSTM1* gene can modify susceptibility to various cancers, including renal cell carcinoma (44,45). Indeed, it has been demonstrated that the carriers of *GSTM1-null* genotype are in significantly higher risk of developing ccRCC compared to the carriers of *GSTM1-active* genotype (60).

GSTT1 deficiency is also a result of the gene deletion. After it has been discovered that GSTT1 could catalyze activation of certain compounds to even more reactive intermediates (28,29,31), the *GSTT1* deletion polymorphism was the subject of many studies, some of which tried to establish whether the presence of the *GSTT1-active* genotype was associated with RCC development, independently or in combination with exposure to certain environmental or occupational hazards (32,61,62). However, the results available in the have shown that *GSTT1* genotype does not, at least independently, affect the susceptibility to RCC (60,63).

The expression of *GSTA1* is exclusively observed in clear cell RCC (58), while in RCC of the chromophobic cell type this protein is completely absent (64). However, the data on the potential role of *GSTA1* SNP in both onset and prognosis of RCC are limited, showing the lack of association in terms of increased risk for RCC development (60,65).

Although the certain meta-analyses on GST polymorphisms in RCC did not report any individual association between *GSTP1* genotype and RCC (63,66), recent results indicated that *GSTP1-variant (ValVal)* genotype was associated with a significant individual risk for ccRCC development, that was even more pronounced in combination with other GST genotypes (60). Namely, if genetic susceptibility to RCC development is, at least partially, affected by polymorphisms in genes involved in xenobiotic metabolism, it is possible that the combinations of certain genotypes may be more discriminating as risk factor for RCC development than a single one. Interestingly, when association of combined GST genotypes was analyzed in terms of RCC risk, *GSTM1-null/GSTT1-active/GSTA1 low-activity/GSTP1-variant* genotype combination was recognized as “the RCC risk carrying genotype”(60).

Glutathione S-transferase (GST), xenobiotic-metabolizing enzymes, play an important role in protection from carcinogens. Presumably, GST genotyping could identify individuals in whom detoxification is diminished, due to complete lack or alteration in enzyme activity. Consequently, they are more likely to accumulate carcinogen-DNA-adducts and/or mutations, increasing their susceptibility to cancer development. Namely, DNA adducts associated with tobacco smoking have been suggested as a marker of biologically effective dose of tobacco carcinogens that might improve individual cancer risk prediction (67). Both free radicals and reactive polycyclic aromatic hydrocarbons metabolites, such as BPDE, are detoxified by GSTs (68,69). Indeed, it has been shown that the clear cell RCC smokers with *GSTM1-null* genotype had significantly higher concentration of BPDE-DNA adducts in comparison with *GSTM1-active* RCC smokers (60).

FUTURE PERSPECTIVES

Some studies suggest that cytosolic GST may be implicated not solely in the development, but also in the progression of RCC (55,70). Although the associations between the certain GST genotypes and RCC risk has been debated in the literature (63,66,70-73), the data on the prognostic value of GST polymorphism in patients with RCC are scarce (55), probably due to the fact that the molecular mechanism supporting the role of GSTs in RCC progression has not been clarified as yet. A possible underlying mechanism might be the regulation of one major signaling pathway, constituting mitogen-activated protein kinases (MAPK) by GSTs. According to the results obtained *in vivo* and *in vitro* setting, mouse *GSTM1-1* physically interacts with ASK1, functioning as ASK1 negative regulator (41). It seems that the ASK1-JNK/p38 pathway is recognized as quite important in the occurrence of the apoptosis in RCC cells (74). Thus, the patients with *GSTM1-null* genotype and consequently deficient *GSTM1*, might have decreased tumor proliferation due to increased apoptotic activity, leading to slower RCC progression and better survival. On the other hand, RCC pa-

tients with *GSTM1-active* genotype may have lower ASK1 activity, resulting in decreased apoptotic activity in the tumor and poorer survival.

Moreover, monomeric GSTP1 subunits inhibit JNK1 by either blocking phosphorylation of JNK or by promoting dephosphorylation of phosphorylated JNK (75). In this manner, JNK is prevented from activating downstream targets in the apoptotic pathway, which might contribute to tumor progression or even drug resistance. Namely, with regard to this role, high tumor GSTP1 expression has been associated with drug resistance, failure of therapy and poor patient survival. Interestingly, GSTP1 overexpression has been found in drug resistant cells, even in instances where there is no evidence that the selecting drug is a substrate for GSTP1 (52).

So far, there are no data which would indicate the significance of *GSTM1:ASK1* and *GSTP1:JNK1* protein: protein interactions in human RCC in terms of tumor progression. What is more, it is still unclear whether the polymorphic expression of *GSTM1* may influence the activity of apoptotic signal pathways in RCC progression.

CONCLUSION

Due to the potential functional significance of common polymorphisms in genes encoding cytosolic glutathione transferase A1, M1, T1 and P1, in both onset and prognosis of RCC, it might be speculated that the presence of specific *GST* gene variants in RCC patients is not only associated with the risk of RCC development, but might also affect the tumor progression and postoperative prognosis.

REFERENCES

- Escudier B, Porta C, Schmidinger M, Algaba F, Patard JJ, Khoo V, et al. Renal cell carcinoma: ESMO Clinical Practice Guidelines for diagnosis, treatment and follow-up. *Ann Oncol.* 2014 Sep 1;25(suppl 3):iii49-iii56.
- Ljungberg B, Bensalah K, Canfield S, Dabestani S, Hofmann F, Hora M, et al. EAU guidelines on renal cell carcinoma: 2014 update. *Eur Urol.* 2015 May;67(5):913-24.
- Patard J-J, Leray E, Rioux-Leclercq N, Cindolo L, Ficarra V, Zisman A, et al. Prognostic value of histologic subtypes in renal cell carcinoma: a multicenter experience. *J Clin Oncol Off J Am Soc Clin Oncol.* 2005 Apr 20;23(12):2763-71.
- Srigley JR, Delahunt B, Eble JN, Egevad L, Epstein JI, Grignon D, et al. The International Society of Urological Pathology (ISUP) Vancouver Classification of Renal Neoplasia. *Am J Surg Pathol.* 2013 Oct;37(10):1469-89.
- Ferlay J, Soerjomataram I, Dikshit R, Eser S, Mathers C, Rebelo M, et al. Cancer incidence and mortality worldwide: sources, methods and major patterns in GLOBOCAN 2012. *Int J Cancer.* 2015 Mar 1;136(5):E359-386.
- Cancer incidence and therapy in central Serbia 2013 [Internet]. Institute of Public Health of Serbia "Dr Milan Jovanović Batut"; 2015. Available from: <http://www.batut.org.rs/download/publikacije/Incidencija%20i%20mortalitet%20od%20raka%202013.pdf>
- Gill IS, Aron M, Gervais DA, Jewett MAS. Clinical practice. Small renal mass. *N Engl J Med.* 2010 Feb 18;362(7):624-34.
- Krabbe L-M, Bagrodia A, Margulis V, Wood CG. Surgical management of renal cell carcinoma. *Semin Interv Radiol.* 2014 Mar;31(1):27-32.
- Ljungberg B, Campbell SC, Cho HY, Jacqmin D, Lee JE, Weikert S, et al. The Epidemiology of Renal Cell Carcinoma. *Eur Urol.* 2011 Oct;60(4):615-21.
- Petejova N, Martinek A. Renal cell carcinoma: Review of etiology, pathophysiology and risk factors. *Biomed Pap.* 2016 Jun 24;160(2):183-94.
- Capitanio U, Montorsi F. Renal cancer. *The Lancet.* 2016 Feb;387(10021):894-906.
- Escudier B, Michaelson MD, Motzer RJ, Hutson TE, Clark JI, Lim HY, et al. Axitinib versus sorafenib in advanced renal cell carcinoma: subanalyses by prior therapy from a randomised phase III trial. *Br J Cancer.* 2014 Jun 10;110(12):2821-8.
- Terris M, Klaassen Z, Kabaria R. Renal cell carcinoma: links and risks. *Int J Nephrol Renov Dis.* 2016 Mar;45.
- International Agency for Research on Cancer, International Agency for Research on Cancer, editors. Beryllium, cadmium, mercury, and exposures in the glass manufacturing industry: ... views and expert opinions of an IARC Working Group on the Evaluation of Carcinogenic Risks to Humans, which met in Lyon, 9 - 16 February 1993. Lyon; 1993. 444 p. (IARC monographs on the evaluation of carcinogenic risks to humans).
- Alexandrov K, Cascorbi I, Rojas M, Bouvier G, Kriek E, Bartsch H. CYP1A1 and GSTM1 genotypes affect benzo[a]pyrene DNA adducts in smokers' lung: comparison with aromatic/hydrophobic adduct formation. *Carcinogenesis.* 2002 Dec;23(12):1969-77.
- Slaga TJ, Bracken WJ, Gleason G, Levin W, Yagi H, Jerina DM, et al. Marked differences in the skin tumor-initiating activities of the optical enantiomers of the diastereomeric benzo(a)pyrene 7,8-diol-9,10-epoxides. *Cancer Res.* 1979 Jan;39(1):67-71.
- Ketterer B. Effects of genetic polymorphism and enzyme induction in the glutathione S-transferase family on chemical safety and risk assessment. *Environ Toxicol Pharmacol.* 1996 Oct 15;2(2-3):157-60.
- Foulkes AS. Genetic Association Studies. In: Applied Statistical Genetics with R [Internet]. New York, NY: Springer New York; 2009 [cited 2016 Jun 28]. p. 1-27. Available from: http://link.springer.com/10.1007/978-0-387-89554-3_1
- Board PG, Menon D. Glutathione transferases, regulators of cellular metabolism and physiology. *Biochim Biophys Acta BBA - Gen Subj.* 2013 May;1830(5):3267-88.
- Hollman A, Tchounwou P, Huang H-C. The Association between Gene-Environment Interactions and Diseases Involving the Human GST Superfamily with SNP Variants. *Int J Environ Res Public Health.* 2016 Mar 29;13(4):379.
- Hayes JD, Flanagan JU, Jowsey IR. Glutathione transferases. *Annu Rev Pharmacol Toxicol.* 2005;45:51-88.
- Laborde E. Glutathione transferases as mediators of signaling pathways involved in cell proliferation and cell death. *Cell Death Differ.* 2010 Sep;17(9):1373-80.
- Grek CL, Zhang J, Manevich Y, Townsend DM, Tew KD. Causes and consequences of cysteine S-glutathionylation. *J Biol Chem.* 2013 Sep 13;288(37):26497-504.
- Wu B, Dong D. Human cytosolic glutathione transferases:

- structure, function, and drug discovery. *Trends Pharmacol Sci.* 2012 Dec;33(12):656–68.
25. Egner PA, Kensler TW, Chen J-G, Gange SJ, Groopman JD, Friesen MD. Quantification of sulforaphane mercapturic acid pathway conjugates in human urine by high-performance liquid chromatography and isotope-dilution tandem mass spectrometry. *Chem Res Toxicol.* 2008 Oct;21(10):1991–6.
 26. Teichert J, Sohr R, Hennig L, Baumann F, Schoppmeyer K, Patzak U, et al. Identification and quantitation of the N-acetyl-L-cysteine S-conjugates of bendamustine and its sulfoxides in human bile after administration of bendamustine hydrochloride. *Drug Metab Dispos Biol Fate Chem.* 2009 Feb;37(2):292–301.
 27. Josephy PD. Genetic variations in human glutathione transferase enzymes: significance for pharmacology and toxicology. *Hum Genomics Proteomics HGP.* 2010;2010:876940.
 28. Guengerich FP. Activation of alkyl halides by glutathione transferases. *Methods Enzymol.* 2005;401:342–53.
 29. Thier R, Brüning T, Roos PH, Rihs H-P, Golka K, Ko Y, et al. Markers of genetic susceptibility in human environmental hygiene and toxicology: the role of selected CYP, NAT and GST genes. *Int J Hyg Environ Health.* 2003 Jun;206(3):149–71.
 30. Kurtovic S, Grehn L, Karlsson A, Hellman U, Mannervik B. Glutathione transferase activity with a novel substrate mimics the activation of the prodrug azathioprine. *Anal Biochem.* 2008 Apr 15;375(2):339–44.
 31. Brüning T, Lammert M, Kempkes M, Thier R, Golka K, Bolt HM. Influence of polymorphisms of GSTM1 and GSTT1 for risk of renal cell cancer in workers with long-term high occupational exposure to trichloroethene. *Arch Toxicol.* 1997;71(9):596–9.
 32. Karami S, Boffetta P, Rothman N, Hung RJ, Stewart T, Zaridze D, et al. Renal cell carcinoma, occupational pesticide exposure and modification by glutathione S-transferase polymorphisms. *Carcinogenesis.* 2008 Jul 1;29(8):1567–71.
 33. Hayes JD, Pulford DJ. The glutathione S-transferase supergene family: regulation of GST and the contribution of the isoenzymes to cancer chemoprotection and drug resistance. *Crit Rev Biochem Mol Biol.* 1995;30(6):445–600.
 34. Inoue T, Irikura D, Okazaki N, Kinugasa S, Matsumura H, Uodome N, et al. Mechanism of metal activation of human hematopoietic prostaglandin D synthase. *Nat Struct Biol.* 2003 Apr;10(4):291–6.
 35. Tars K, Olin B, Mannervik B. Structural basis for featuring of steroid isomerase activity in alpha class glutathione transferases. *J Mol Biol.* 2010 Mar 19;397(1):332–40.
 36. Hayes JD, McLellan LI. Glutathione and glutathione-dependent enzymes represent a co-ordinately regulated defence against oxidative stress. *Free Radic Res.* 1999 Oct;31(4):273–300.
 37. Seeley SK, Poposki JA, Maksimchuk J, Tebbe J, Gaudreau J, Mannervik B, et al. Metabolism of oxidized linoleic acid by glutathione transferases: peroxidase activity toward 13-hydroperoxyoctadecadienoic acid. *Biochim Biophys Acta.* 2006 Jul;1760(7):1064–70.
 38. McIlwain CC, Townsend DM, Tew KD. Glutathione S-transferase polymorphisms: cancer incidence and therapy. *Oncogene.* 2006 Mar 13;25(11):1639–48.
 39. Tew KD, Townsend DM. Glutathione-s-transferases as determinants of cell survival and death. *Antioxid Redox Signal.* 2012 Dec 15;17(12):1728–37.
 40. Adler V, Yin Z, Fuchs SY, Benezra M, Rosario L, Tew KD, et al. Regulation of JNK signaling by GSTp. *EMBO J.* 1999 Mar 1;18(5):1321–34.
 41. Cho SG, Lee YH, Park HS, Ryoo K, Kang KW, Park J, et al. Glutathione S-transferase mu modulates the stress-activated signals by suppressing apoptosis signal-regulating kinase 1. *J Biol Chem.* 2001 Apr 20;276(16):12749–55.
 42. Ichijo H, Nishida E, Irie K, ten Dijke P, Saitoh M, Moriguchi T, et al. Induction of apoptosis by ASK1, a mammalian MAPKKK that activates SAPK/JNK and p38 signaling pathways. *Science.* 1997 Jan 3;275(5296):90–4.
 43. Dorion S, Lambert H, Landry J. Activation of the p38 signaling pathway by heat shock involves the dissociation of glutathione S-transferase Mu from Ask1. *J Biol Chem.* 2002 Aug 23;277(34):30792–7.
 44. Hayes JD, Strange RC. Glutathione S-transferase polymorphisms and their biological consequences. *Pharmacology.* 2000 Sep;61(3):154–66.
 45. Di Pietro G, Magno LAV, Rios-Santos F. Glutathione S-transferases: an overview in cancer research. *Expert Opin Drug Metab Toxicol.* 2010 Feb;6(2):153–70.
 46. Guengerich FP, McCormick WA, Wheeler JB. Analysis of the kinetic mechanism of haloalkane conjugation by mammalian theta-class glutathione transferases. *Chem Res Toxicol.* 2003 Nov;16(11):1493–9.
 47. Coles BF, Kadlubar FF. Human alpha class glutathione S-transferases: genetic polymorphism, expression, and susceptibility to disease. *Methods Enzymol.* 2005;401:9–42.
 48. Coles BF, Kadlubar FF. Detoxification of electrophilic compounds by glutathione S-transferase catalysis: determinants of individual response to chemical carcinogens and chemotherapeutic drugs? *BioFactors Oxf Engl.* 2003;17(1–4):115–30.
 49. Dusinská M, Ficek A, Horská A, Raslová K, Petrovská H, Vallová B, et al. Glutathione S-transferase polymorphisms influence the level of oxidative DNA damage and antioxidant protection in humans. *Mutat Res.* 2001 Oct 1;482(1–2):47–55.
 50. Hu X, O'Donnell R, Srivastava SK, Xia H, Zimniak P, Nanduri B, et al. Active site architecture of polymorphic forms of human glutathione S-transferase P1-1 accounts for their enantioselectivity and disparate activity in the glutathione conjugation of 7beta,8alpha-dihydroxy-9alpha,10alpha-ox y-7,8,9,10-tetrahydrobenzo(a)pyrene. *Biochem Biophys Res Commun.* 1997 Jun 18;235(2):424–8.
 51. Townsend DM, Findlay VL, Tew KD. Glutathione S-transferases as regulators of kinase pathways and anticancer drug targets. *Methods Enzymol.* 2005;401:287–307.
 52. Townsend DM, Tew KD. The role of glutathione-S-transferase in anti-cancer drug resistance. *Oncogene.* 2003 Oct 20;22(47):7369–75.
 53. Reinemer P, Dirr HW, Ladenstein R, Huber R, Lo Bello M, Federici G, et al. Three-dimensional structure of class pi glutathione S-transferase from human placenta in complex with S-hexylglutathione at 2.8 Å resolution. *J Mol Biol.* 1992 Sep 5;227(1):214–26.
 54. Ahmad ST, Arjumand W, Seth A, Kumar Saini A, Sultana S. Impact of glutathione transferase M1, T1, and P1 gene polymorphisms in the genetic susceptibility of North Indian population to renal cell carcinoma. *DNA Cell Biol.* 2012 Apr;31(4):636–43.

55. De Martino M, Klatte T, Schatzl G, Remzi M, Waldert M, Haitel A, et al. Renal cell carcinoma Fuhrman grade and histological subtype correlate with complete polymorphic deletion of glutathione S-transferase M1 gene. *J Urol*. 2010 Mar;183(3):878–83.
56. Salinas-Sánchez AS, Sánchez-Sánchez F, Donate-Moreno MJ, Rubio-del-Campo A, Serrano-Oviedo L, Gimenez-Bachs JM, et al. GSTT1, GSTM1, and CYP1B1 gene polymorphisms and susceptibility to sporadic renal cell cancer. *Urol Oncol Semin Orig Investig*. 2012 Nov;30(6):864–70.
57. Sweeney C, Farrow DC, Schwartz SM, Eaton DL, Checkoway H, Vaughan TL. Glutathione S-transferase M1, T1, and P1 polymorphisms as risk factors for renal cell carcinoma: a case-control study. *Cancer Epidemiol Biomark Prev Publ Am Assoc Cancer Res Cosponsored Am Soc Prev Oncol*. 2000 Apr;9(4):449–54.
58. Simic T, Savic-Radojevic A, Pljesa-Ercegovac M, Matic M, Mimic-Oka J. Glutathione S-transferases in kidney and urinary bladder tumors. *Nat Rev Urol*. 2009 May;6(5):281–9.
59. Green T, Dow J, Ellis MK, Foster JR, Odum J. The role of glutathione conjugation in the development of kidney tumours in rats exposed to trichloroethylene. *Chem Biol Interact*. 1997 Jul 11;105(2):99–117.
60. Coric VM, Simic TP, Pekmezovic TD, Basta-Jovanovic GM, Savic Radojevic AR, Radojevic-Skodric SM, et al. Combined GSTM1-Null, GSTT1-Active, GSTA1 Low-Activity and GSTP1-Variant Genotype Is Associated with Increased Risk of Clear Cell Renal Cell Carcinoma. *PLoS One*. 2016;11(8):e0160570.
61. Buzio L, De Palma G, Mozzoni P, Tondel M, Buzio C, Franchini I, et al. Glutathione S-transferases M1-1 and T1-1 as risk modifiers for renal cell cancer associated with occupational exposure to chemicals. *Occup Environ Med*. 2003 Oct;60(10):789–93.
62. Longuemaux S, Deloménie C, Gallou C, Méjean A, Vincent-Viry M, Bouvier R, et al. Candidate genetic modifiers of individual susceptibility to renal cell carcinoma: a study of polymorphic human xenobiotic-metabolizing enzymes. *Cancer Res*. 1999 Jun 15;59(12):2903–8.
63. Yang X, Long S, Deng J, Deng T, Gong Z, Hao P. Glutathione S-Transferase Polymorphisms (GSTM1, GSTT1 and GSTP1) and Their Susceptibility to Renal Cell Carcinoma: An Evidence-Based Meta-Analysis. Medeiros R, editor. *PLoS ONE*. 2013 May 22;8(5):e63827.
64. Liu L, Qian J, Singh H, Meiers I, Zhou X, Bostwick DG. Immunohistochemical analysis of chromophobe renal cell carcinoma, renal oncocytoma, and clear cell carcinoma: an optimal and practical panel for differential diagnosis. *Arch Pathol Lab Med*. 2007 Aug;131(8):1290–7.
65. Searchfield L, Price SA, Betton G, Jasani B, Riccardi D, Griffiths DFR. Glutathione S-transferases as molecular markers of tumour progression and prognosis in renal cell carcinoma: GST-alpha in human RCC. *Histopathology*. 2011 Jan;58(2):180–90.
66. Jia C-Y, Liu Y-J, Cong X-L, Ma Y-S, Sun R, Fu D, et al. Association of glutathione S-transferase M1, T1, and P1 polymorphisms with renal cell carcinoma: evidence from 11 studies. *Tumor Biol*. 2014 Apr;35(4):3867–73.
67. Wiencke JK. DNA adduct burden and tobacco carcinogenesis. *Oncogene*. 2002 Oct 21;21(48):7376–91.
68. Filiadis I, Hrouda D. Genetic factors in chemically-induced transitional cell bladder cancer. *BJU Int*. 2000 Nov;86(7):794–801.
69. Jung I, Messing E. Molecular mechanisms and pathways in bladder cancer development and progression. *Cancer Control J Moffitt Cancer Cent*. 2000 Aug;7(4):325–34.
70. Huang W, Shi H, Hou Q, Mo Z, Xie X. GSTM1 and GSTT1 polymorphisms contribute to renal cell carcinoma risk: evidence from an updated meta-analysis. *Sci Rep*. 2015;5:17971.
71. Abid A, Ajaz S, Khan AR, Zehra F, Hasan AS, Sultan G, et al. Analysis of the glutathione S-transferase genes polymorphisms in the risk and prognosis of renal cell carcinomas. Case-control and meta-analysis. *Urol Oncol Semin Orig Investig [Internet]*. 2016 May [cited 2016 May 14]; Available from: <http://linkinghub.elsevier.com/retrieve/pii/S1078143916300151>
72. Cheng H-Y, You H-Y, Zhou T-B. Relationship between GSTM1/GSTT1 Null Genotypes and Renal Cell Carcinoma Risk: A Meta-Analysis. *Ren Fail*. 2012 Sep;34(8):1052–7.
73. Liu R, Wang X-H, Liu L, Zhou Q. No association between the GSTM1 null genotype and risk of renal cell carcinoma: a meta-analysis. *Asian Pac J Cancer Prev APJCP*. 2012;13(7):3109–12.
74. Hassan M, Feyen O, Grinstein E. Fas-induced apoptosis of renal cell carcinoma is mediated by apoptosis signal-regulating kinase 1 via mitochondrial damage-dependent caspase-8 activation. *Cell Oncol Off J Int Soc Cell Oncol*. 2009;31(6):437–56.
75. Adler V, Pincus MR. Effector peptides from glutathione-S-transferase-pi affect the activation of jun by jun-N-terminal kinase. *Ann Clin Lab Sci*. 2004;34(1):35–46.