

## REVIEW PAPER

UDK: 615.363:615.322

DOI: 10.5937/hralsh2202001C

# Phytoestrogens as estrogen receptor ligands

Aleksandra Cvoro

Faculty of Biology, University of Belgrade,  
Studentski trg 16, Belgrade, Serbia

Corresponding author:

Dr Aleksandra Cvoro

Faculty of Biology, University of Belgrade  
Studentski trg 16, 11000 Belgrade, Serbia

E-mail address: aleksandra.cvoro@bio.bg.ac.rs

Received 01 September 2023

Accepted 22 October 2023

**Abstract**

*Estrogen receptors (ERs) are ligand-regulated transcription factors that modulate essential transcriptional programs by either promoting or repressing targeted gene expression. Given the impact of ER signaling on development, metabolism and physiology, it is no surprise to find impaired ER function as the basis of many disorders; thus, ERs have long been recognized as important biological and pharmaceutical targets. While 17 $\beta$ -estradiol (E2) is the main ER cognate ligand, ERs can be activated by diverse estrogen-mimicking compounds e.g. phytoestrogens, capable of binding receptors in a variable manner and influencing estrogen-dependent pathways, with both, beneficial and harmful health consequences. In this review we assessed current knowledge in the field of phytoestrogens as ER alternative ligands.*

**Key words:** Estrogen receptor; phytoestrogens; MF101; genistein.

**INTRODUCTION**

Estrogen receptors (ERs) are ligand-dependent transcription factors that facilitate the regulation of numerous physiological programs such as development, metabolism and homeostasis. The predominant endogenous ligand for ERs is estrogenic compound E2. It is synthesized and secreted mainly by the ovaries in females, and testes and adrenal cortex in males. Nevertheless, E2 is produced in a number of other tissues including adipose tissue, liver, skin, pancreas and brain [1,2,3]. Since ERs are expressed in numerous tissues, estrogens exhibit complex multisystemic effects, and are, consequently, implicated in the development and progression of various diseases.

While E2 is the natural ligand for ERs, these receptors can also be activated by various endogenous and exogenous ligands capable of binding ERs and inducing diverse receptor conformations that affect estrogen-dependent gene regulation. These ER ligands are acting by promoting (i.e., agonists) or blocking estrogenic activity (i.e., antagonists), modifying a variety of biological effects. Understanding agonistic/antagonistic features of a ligand demands the recognition of ligands interference with receptor activity at various levels: from the stability of ER-transcriptional complex, through ER's homo- and heterodimerization

ability to its interaction with DNA response elements. Since ligand characteristics are highly diverse, a small difference in a structure can cause critical difference in outcome and add intricacy to overall ER-mediated effects. The most famous examples are selective estrogen receptor modulators (SERMs) that are targeting the ER for the treatment of endocrine disorders [4,5]. Although SERMs bind the same site on ER as E2 they exert selective agonist or antagonist activity in different target tissues as a consequence of unique SERM-receptor conformations.

A distinctive group of ER ligands are phytoestrogens, a form of dietary plant-derived estrogen-mimicking compounds. Phytoestrogens could affect the function of estrogens in different ways. They can modulate estrogen synthesis and metabolism or transport of the estrogens throughout the body [6,7]. But the best examined phytoestrogens are those acting as ER ligands i.e., binding directly to ERs and competing with endogenous estrogens [8]. The overall effect of phytoestrogens will depend upon sex, age and health status of the consumer, as well as dose, type and synergistic interactions among consumed compounds, the level of endogenous estrogens and ER isoforms presence [9,10,11].

## ESTROGEN RECEPTORS: STRUCTURE AND MECHANISMS OF ACTION

Most estrogen hormone effects are mediated by two different ERs, ER $\alpha$  and ER $\beta$  [12,13]. ER $\alpha$  and ER $\beta$  are encoded by different genes; the human ER $\alpha$  gene is located on chromosome 6q25.1 [14], whereas the human ER $\beta$  gene is located on chromosome 14q23.2 [15]. ER $\alpha$  and ER $\beta$  exert disparate biological roles, as evidenced by their distinct intracellular and tissue distribution patterns [16,17], different gene regulation patterns [18] and different phenotypes detected in ER $\alpha$  and ER $\beta$  knockout mice [19].

As members of the nuclear receptor superfamily [20], ER $\alpha$  and ER $\beta$  have a general multidomain structure. The ERs consist of six functional domains: a variable, N-terminal A/B domain which contains activation function-1 (AF-1) that can activate gene transcription in a constitutive manner; a central C domain known as the DNA-binding domain (DBD) responsible for binding to specific DNA sequences known as estrogen response elements (ERE); linker or hinge region between the LBD and DBD known as D domain; and a C-terminal E/F domain, which contains the ligand-binding domain (LBD), dimerization interfaces and activation function-2 (AF-2), that serves as a binding site for coregulatory proteins [21]. While DBD in ER $\alpha$  and ER $\beta$  is highly conserved exhibiting 95% homology, the LBD is flexible, with only partial homology (55%).

ERs regulate physiological processes through two major mechanisms. In the conventional genomic pathway of estrogen action, multistep process responsible for ER-mediated gene regulation takes place [22,23]. This mode of action begins by binding of the ligand to the ER LBD. This event produces a conformational change in ER that depends strongly on the nature of the ligand and leads to the receptor dimerization, and binding to the EREs [24]. The ligand binding also induces a conformational change within the ER LBD allowing coregulatory proteins to be recruited [25,26] which culminates in the modulation of target gene expression in response to estrogens. Nevertheless, ERs can regulate transcription without binding directly to DNA. ERs in such cases are acting through protein-protein interactions with a transcription factor complex that contacts the DNA [27].

The second pathway of ER action is the non-genomic mechanism which mediates rapid signaling events [28,29]. In this pathway, initiated at the cell membrane, estrogens bind to membrane-associated ERs - ER $\alpha$ , ER $\beta$  or G protein-coupled estrogen receptor (GPER1), and cause non-genomic effects. These actions include the mobilization of intracellular calcium [30], the stimulation of adenylate cyclase activity and cAMP production [31,32] and the activation of various protein-kinase cascades [33].

It is important to recognize that different mechanisms of ER actions are co-dependent, overlapping and even converging at same target genes. Namely, non-genomic pathway of ER action may indirectly regulate gene expression, through activation of signaling pathways that act on target transcription factors.

## INTERACTION OF PHYTOESTROGENS WITH ERs

Phytoestrogens are plant-derived substances with estrogenic properties. Although they have been used in traditional medicine for a long time, the interest in phytoestrogens is intensified due to the increased amount of information on their effects on human health, as well as better understanding of estrogen action in general. As a consequence of their estrogenic activity, phytoestrogens have been ascribed beneficial health effects such as the alleviation of menopausal symptoms, prevention of osteoporosis, reduced risks of cardiovascular disease, obesity, and even breast, endometrial and prostate cancer. However, adverse effects of phytoestrogens have also been reported [34]. As already mentioned, majority of research in phytoestrogen signaling has focused on their role as ER ligands, i.e. their ability and the ability of their metabolites to interact and activate ERs.

Phytoestrogens are highly diverse group found in more than 300 plant species [35] including a wide variety of human foods such as soy products, legume seeds and dairy products [36,37]. According to their chemical structure phytoestrogens are nonsteroidal polyphenols. Their structural similarity with estrogens (e.g., the presence of phenolic rings) is a requirement for phytoestrogen to bind ER. Essentially, they are classified in two groups: flavonoids and non-flavonoids [38]. Flavonoids have the fifteen-carbon skeleton (C6-C3-C6) which consists of two benzene rings (A and B) and a heterocyclic pyran structure (C). Best characterized groups of flavonoids are isoflavones, coumestans and prenylflavonoids (**Figure 1**). This classification is based on structural differences [39] and modifications in the skeleton and/or the chemical groups of flavonoids [Reviewed in detail in 40]. Non-flavonoids consist of naturally occurring phenolic acids, either hydroxybenzoic acid (C6-C1 structure) or hydroxycinnamic acid (C6-C3 structure) [39]. Main representatives of non-flavonoid phytoestrogens are lignans and stilbenes (**Figure 2**).

In plants, the phytoestrogens perform mainly a protective and defense functions exerting anti-parasitic, antibacterial, antiviral and fungistatic effects. They also act as antioxidants, protecting the plant from harmful effects of sunlight [41].

In the human body, the phytoestrogens are rarely active as free molecules. They are metabolized both by tissue enzymes and gastrointestinal microbiota, either

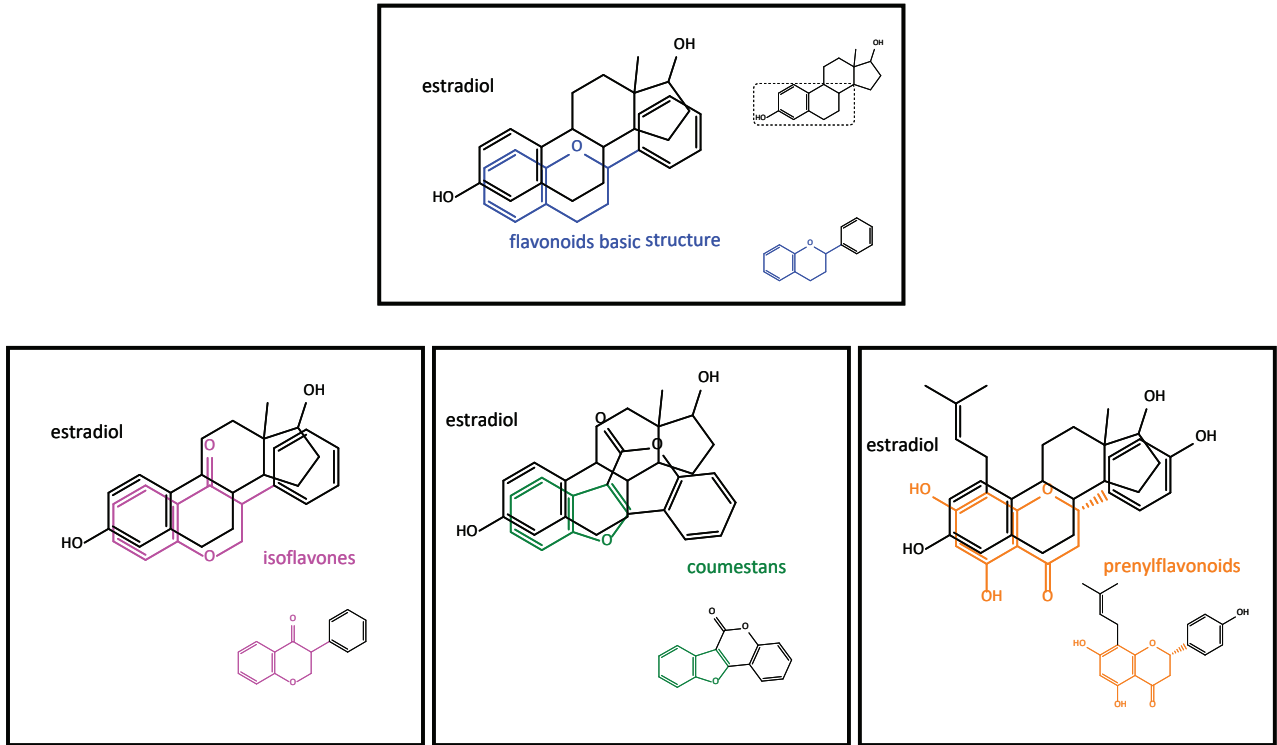


Figure 1. Structural similarity between estradiol and major flavonoids.

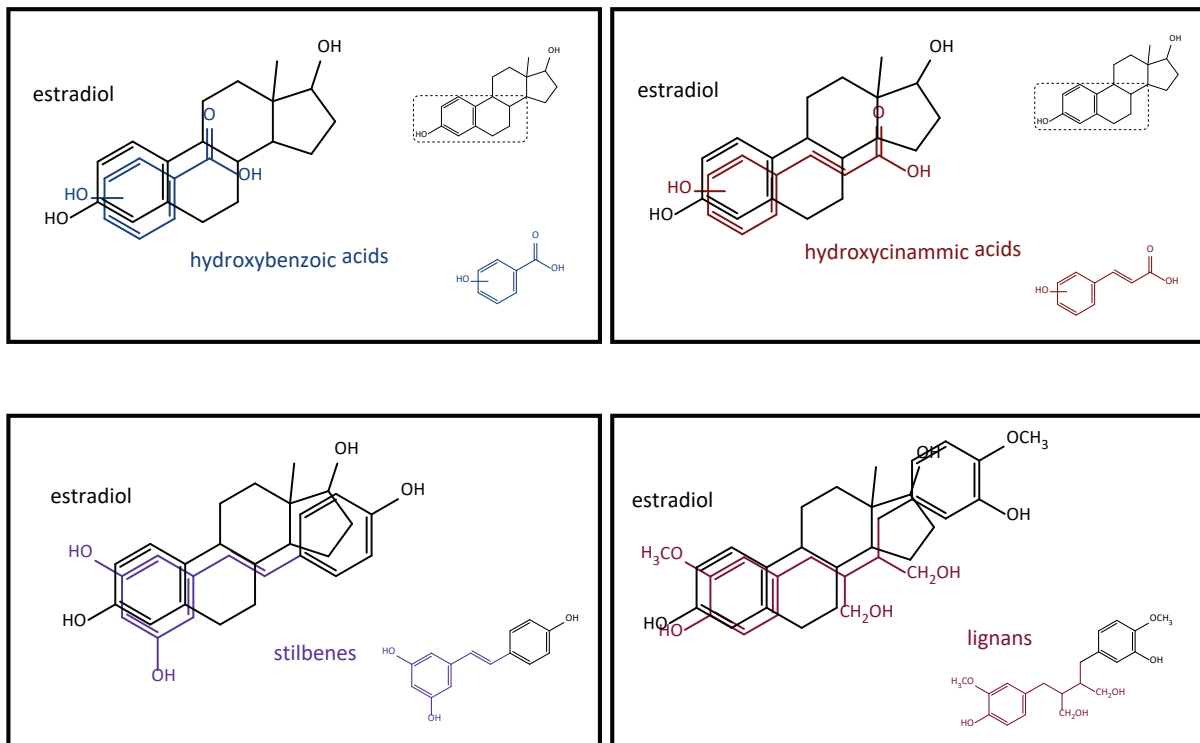


Figure 2. Structural similarity between estradiol and major non-flavonoids.

prior to absorption or during enterohepatic circulation. Notably, individual phytoestrogen metabolites have often higher ER binding affinity than phytoestrogen itself. Most of ingested phytoestrogens are unabsorbed in the proximal intestine [42,43] but reach

the colon where they are exposed to microbiota and undergo metabolization [44,45]. Gut microbiota is assumed to play an essential role in the metabolism of phytoestrogens in humans by impacting the degree to which they are absorbed, metabolized and degraded.

Thus, the differences in an individual's distinctive gut microbiota, which is determined by many factors including diet, bowel disease, stress, gender, age and genetics, may play an important role in the bioavailability of phytoestrogens and their metabolites, and the outcomes of any subsequent biological response.

### MOLECULAR MECHANISMS OF PHYTOESTROGEN ACTION

Since the majority of phytoestrogen effects are mediated through ERs, it is evident that small differences in phytoestrogen/ER structural conformation will cause significant differences in phytoestrogen biological activity. Thus, the difference between the actions of particular phytoestrogens can be explained in large part by differences in their ability to influence ER transcriptional functions.

The transcriptional, and consequently physiological response to a phytoestrogen depends on a number of factors, including: 1) differences in ER subtype affinity and selectivity, 2) differences in recruitment of coregulatory proteins and /or 3) differences in promoter context, i.e., altered specificity and affinity of the DNA response elements through which the ER is acting. As a result, phytoestrogens have a high potential for modifying all estrogen-dependent biological activities and cell signaling pathways in all imaginable ways [Reviewed in detail in 40].

One of the most extensively studied phytoestrogens is the isoflavone genistein that corresponds to the chemical name '4',5,7-Trihydroxyisoflavone'. It is typically found in legumes, such as soybeans and fava beans, and is being metabolized by endogenous phase I and phase II enzymes as well as by the gut bacteria leading to production of metabolites with altered bioactivity [46,47]. Majority of its effects at nutritionally relevant concentrations are ER-mediated [48]. Furthermore, genistein predominantly employs ER classical genomic mechanism [49].

Insight into the genistein mechanism of action is connected with understanding the differences in ER $\alpha$  vs ER $\beta$  action (**Table 1**). Although ER $\alpha$  and ER $\beta$  are mediating multiple biological effects in a similar manner, their actions are sometimes resulting in entirely opposing effects. Namely, ERs are modulators of cell growth and differentiation but ER $\alpha$  stimulates cell growth and proliferation, while ER $\beta$  is associated with cell differentiation and tumor suppression [59]. On the other hand, both ER $\alpha$  and ER $\beta$  are equally effective in transcriptional repression of inflammatory genes responsible for osteoporosis [60] and other beneficial ER effects. Consequently, much effort has been applied in developing and/or discovering ER $\alpha$  and ER $\beta$  selective ligands that will have selective clinical effects and uncouple beneficial actions of ERs from deleterious effects.

**Table 1.** ER subtype specificity of representative phytoestrogens.

Name	Class	ER specificity	Reference
<b>FLAVO-NOIDS</b>			
genistein	isoflavones	ER $\beta$ > ER $\alpha$	[50]
daidzein	isoflavones	ER $\beta$ > ER $\alpha$	[49,51]
biochanin A	isoflavones	ER $\beta$ > ER $\alpha$	[52]
quercetin	flavonol	ER $\beta$ > ER $\alpha$	[53-55]
coumestrol	coumestans	ER $\beta$ > ER $\alpha$	[50]
8-prenylnar-ingenin	prenylflavonoids	ER $\alpha$ $\geq$ ER $\beta$	[56,57]
<b>NON-FLAVO-NOIDS</b>			
enterolactone	lignans	ER $\alpha$ > ER $\beta$	[50,53,58]
resveratrol	stilbenes	ER $\alpha$ > ER $\beta$	[50,53]

Genistein, like most of the phytoestrogens identified so far, is ER $\beta$ -selective, and as such may act as natural SERM [61]. The genistein selectiveness is direct consequence of difference in genistein relative binding affinity given that genistein binds with a 20-fold greater affinity for ER $\beta$  than ER $\alpha$  [49]. Thus, genistein offers a great potential as ER $\beta$  selective ligand, which is avoiding cancer-promoting effects of ER $\alpha$ , but keeping beneficial ER $\beta$ -mediated effects.

On the molecular level, genistein modulates various steps of cell cycle through ER, in order to induce the desired antiproliferative effect. The regulation of cell proliferation by genistein implores multiple mechanisms leading to G2/M cell cycle arrest. These mechanisms are primarily driven by the regulation of crucial cell cycle regulatory genes including several cyclins, among them cyclin-dependent kinases (CDKs) and cyclin-dependent kinase inhibitors (CDKIs). Another genistein-mediated mechanism engaged in G2/M cell cycle arrest involves the Ras/MAPK/activator protein-1 signaling pathway and results in the suppression of the transcription factors Cdk1, cyclin B1, and Cdc25C [62].

In other context and as a consequence of multiple levels of action, genistein has antioxidant effect. There is direct antioxidant effect of genistein that derives from its capacity to act as a ROS scavenger [63,64]. Furthermore, genistein through ER stimulates the expression of the antioxidant enzymes manganese superoxide dismutase, glutathione peroxidase and catalase [65,66].

However, it will be way too simple to consider the effects of genistein (or any other phytoestrogen or ER ligand in general) exclusively in the light of cell cycle regulation and/or antioxidant effects. Multiple mechanisms producing antiangiogenic, antimetastatic, anti-inflammatory and anti-amyloid- $\beta$  activities are not

going to be considered here [67,68]. Besides, most of our knowledge about phytoestrogens is the result of the studies concentrated on one compound at a time, extremely unlikely context to be found in reality.

In fact, we are constantly exposed to a myriad of compounds that are competing for ERs and producing wide range of effects depending on the fine balance between presence of ERs in tissues, concentrations of endogenous estrogens and the content and dosages of consumed phytoestrogens. Above all, it is necessary to acknowledge synergistic interactions among estrogenic compounds that are as significant as the effects of each of them separately. Brilliant example is MF101 (Menerba) - the herbal extract from number of plants, with distinctive estrogenic activity. MF101 was studied as a potential treatment for hot flashes in postmenopausal women. MF101 formula was based on the use of botanical supplements exploited in Traditional Chinese Medicine for the treatment of menopausal symptoms [69].

What makes it interesting, MF101 is crude herbal extract containing 22 individual plants and multitude of the compounds that synergistically act in order to produce unique effect [70,71]. Despite such a complexity MF101 is ER $\beta$  selective. Unlike genistein and most of the phytoestrogens whose receptor subtype selectivity is a consequence of difference in binding affinity, MF101 binds to both ER $\alpha$  and ER $\beta$  with equal affinity [70]. In this case, ER $\beta$  selectivity of MF101 is due to the selective recruitment of cofactors in respect to difference in conformational change produced by MF101 in ER $\alpha$  vs ER $\beta$  [70].

Thus, MF101-mediated physiological effects such as alleviation of menopausal symptoms like hot flashes [72] with no induction of proliferative events are likely the consequence of synergistical action of multiple compounds. We also shouldn't forget that prescription drugs are often used alongside either phytoestrogen-containing food or herbal supplements with a hope of synergistic beneficial effects but with a chance of unwanted competitive interactions.

## CONCLUSIONS

Recent advances in our understanding of the phytoestrogens as effective ER modulators have spurred substantial interest in their potential to serve as therapeutic agents. Initial studies focused on the conformational binding effects of various ligands that would be beneficial for targeting ERs. However, the standard picture of the particular compound that has beneficial effect on particular condition turned out to be oversimplified. Multiple factors converge on the ER transcriptional regulation to determine the final response. The obvious determinants are variability in dietary phytoestrogen intake: frequency, timing and composition

of consumed phytoestrogens, as well as substantial inter-individual differences in the metabolic and pharmacokinetic processing of ingested phytoestrogens. As already underlined, the presence and distribution of ER subtypes and transcriptional cofactors in different tissues and cell types are adding to complexity of the outcome. Finally, the amount and type of endogenous hormone which will compete for the binding of the ERs with exogenous ligands will determine the overall effect. Nevertheless, such interplay allows ERs to respond dynamically to any ligand.

So, in spite of the extraordinary expansion of the phytoestrogen studies in the last decades, the level of complexity and context-dependent nature of its actions are a real challenge for the future.

## ACKNOWLEDGEMENTS

This work was supported by Ministry of Science, Technological Development and Innovations of the Republic of Serbia (Grant number 451-03-66/2024-03/200178). We thank Prof. Dr. Deana Andric for her invaluable technical assistance and helpful comments on the manuscript.

## REFERENCES

1. Simpson E, Rubin G, Clyne C. Local estrogen biosynthesis in males and females. *Endocr Relat Cancer*. 1999;6:131–137. doi: 10.1677/erc.0.0060131.
2. Nelson LR, Bulun SE. Estrogen production and action. *J Am Acad Dermatol*. 2001;45:5116–5124. doi: 10.1067/mjd.2001.117432.
3. Simpson ER. Sources of estrogen and their importance. *J Steroid Biochem Mol Biol*. 2003;86:225–230. doi: 10.1016/S0960-0760(03)00360-1.
4. Kharode Y, Bodine PV, Miller CP, Lyttle CR, Komm BS. The pairing of a selective estrogen receptor modulator, bazedoxifene, with conjugated estrogens as a new paradigm for the treatment of menopausal symptoms and osteoporosis prevention. *Endocrinology*. 2008;149(12):6084–91. doi: 10.1210/en.2008–0817.
5. Maximov PY, Lee TM, Jordan VC. The discovery and development of selective estrogen receptor modulators (SERMs) for clinical practice. *Curr Clin Pharmacol*. 2013;8(2):135–55. doi: 10.2174/1574884711308020006.
6. van Duursen MBM. Modulation of estrogen synthesis and metabolism by phytoestrogens *in vitro* and the implications for women's health. *Toxicol Res (Camb)*. 2017;6(6):772–794. doi: 10.1039/c7tx00184c.
7. Whitten PL, Patisaul HB. Cross-species and interassay comparisons of phytoestrogen action. *Environ Health Perspect*. 2001;109 Suppl 1(Suppl 1):5–20. doi: 10.1289/ehp.011109s15.
8. Leitman DC, Paruthiyil S, Vivar OI, Saunier EF, Herber CB, Cohen I, Tagliaferri M, Speed TP. Regulation of specific target genes and biological responses by estrogen receptor subtype agonists. *Curr Opin Pharmacol*. 2010;10(6):629–36. doi: 10.1016/j.coph.2010.09.009.

9. Bennetau-Pelissero C. Risks and benefits of phytoestrogens: where are we now? *Curr Opin Clin Nutr Metab Care.* 2016;19(6):477–483. doi: 10.1097/MCO.0000000000000326.
10. Rietjens IMCM, Lousse J, Beekmann K. The potential health effects of dietary phytoestrogens. *Br J Pharmacol.* 2017;174(11):1263–1280. doi: 10.1111/bph.13622.
11. Patisaul HB, Jefferson W. The pros and cons of phytoestrogens. *Front Neuroendocrinol.* 2010;31(4):400–19. doi: 10.1016/j.yfrne.2010.03.003.
12. Chambon P. The nuclear receptor superfamily: a personal retrospect on the first two decades. *Mol Endocrinol.* 2005;19(6):1418–28. doi: 10.1210/me.2005-0125.
13. Koehler KF, Helguero LA, Haldosén LA, Warner M, Gustafsson JA. Reflections on the discovery and significance of estrogen receptor beta. *Endocr Rev.* 2005;26(3):465–78. doi: 10.1210/er.2004-0027.
14. Menasce LP, White GR, Harrison CJ, Boyle JM. Localization of the estrogen receptor locus (ESR) to chromosome 6q25.1 by FISH and a simple post-FISH banding technique. *Genomics.* 1993;17(1):263–5. doi: 10.1006/geno.1993.1320.
15. Enmark E, Peltö-Huikko M, Grandien K, Lagercrantz S, Lagercrantz J, Fried G, Nordenskjöld M, Gustafsson JA. Human estrogen receptor beta-gene structure, chromosomal localization, and expression pattern. *J Clin Endocrinol Metab.* 1997;82(12):4258–65. doi: 10.1210/jcem.82.12.4470.
16. Mueller SO, Korach KS. Estrogen receptors and endocrine diseases: lessons from estrogen receptor knockout mice. *Curr Opin Pharmacol.* 2001;1(6):613–9. doi: 10.1016/S1471-4892(01)00105-9.
17. Velickovic K, Cvoro A, Srdic B, Stokic E, Markelic M, Golic I, Otasevic V, Stancic A, Jankovic A, Vucetic M, Buzadzic B, Korac B, Korac A. Expression and subcellular localization of estrogen receptors  $\alpha$  and  $\beta$  in human fetal brown adipose tissue. *J Clin Endocrinol Metab.* 2014;99(1):151–9. doi: 10.1210/jc.2013-2017.
18. Tee MK, Rogatsky I, Tzagarakis-Foster C, Cvoro A, An J, Christy RJ, Yamamoto KR, Leitman DC. Estradiol and selective estrogen receptor modulators differentially regulate target genes with estrogen receptors alpha and beta. *Mol Biol Cell.* 2004;15(3):1262–72. doi: 10.1091/mbc.e03-06-0360.
19. Hewitt SC, Harrell JC, Korach KS. Lessons in estrogen biology from knockout and transgenic animals. *Annu Rev Physiol.* 2005;67:285–308. doi: 10.1146/annurev.physiol.67.040403.115914.
20. Evans RM. The steroid and thyroid hormone receptor superfamily. *Science.* 1988;240(4854):889–95. doi: 10.1126/science.3283939.
21. Feng W, Ribeiro RC, Wagner RL, Nguyen H, Apriletti JW, Fletterick RJ, Baxter JD, Kushner PJ, West BL. Hormone-dependent coactivator binding to a hydrophobic cleft on nuclear receptors. *Science.* 1998;280(5370):1747–9. doi: 10.1126/science.280.5370.1747.
22. O'Malley BW. A life-long search for the molecular pathways of steroid hormone action. *Mol Endocrinol.* 2005;19(6):1402–11. doi: 10.1210/me.2004-0480.
23. Fuentes N, Silveyra P. Estrogen receptor signaling mechanisms. *Adv Protein Chem Struct Biol.* 2019;116:135–170. doi: 10.1016/bs.apcsb.2019.01.001.
24. Nilsson S, Mäkelä S, Treuter E, Tujague M, Thomsen J, Andersson G, Enmark E, Pettersson K, Warner M, Gustafsson JA. Mechanisms of estrogen action. *Physiol Rev.* 2001;81(4):1535–65. doi: 10.1152/physrev.2001.81.4.1535.
25. Rosenfeld MG, Glass CK. Coregulator codes of transcriptional regulation by nuclear receptors. *J Biol Chem.* 2001;276(40):36865–8. doi: 10.1074/jbc.R100041200.
26. Lonard DM, Lanz RB, O'Malley BW. Nuclear receptor coregulators and human disease. *Endocr Rev.* 2007;28(5):575–87. doi: 10.1210/er.2007-0012.
27. Göttlicher M, Heck S, Herrlich P. Transcriptional cross-talk, the second mode of steroid hormone receptor action. *J Mol Med (Berl).* 1998;76(7):480–9. doi: 10.1007/s001090050242.
28. Hammes SR, Levin ER. Minireview: Recent advances in extranuclear steroid receptor actions. *Endocrinology.* 2011;152(12):4489–95. doi: 10.1210/en.2011-1470.
29. Hammes SR, Levin ER. Extranuclear steroid receptors: nature and actions. *Endocr Rev.* 2007;28(7):726–41. doi: 10.1210/er.2007-0022.
30. Improta-Brears T, Whorton AR, Codazzi F, York JD, Meyer T, McDonnell DP. Estrogen-induced activation of mitogen-activated protein kinase requires mobilization of intracellular calcium. *Proc Natl Acad Sci U S A.* 1999;96(8):4686–91. doi: 10.1073/pnas.96.8.4686.
31. Aronica SM, Kraus WL, Katzenellenbogen BS. Estrogen action via the cAMP signaling pathway: stimulation of adenylate cyclase and cAMP-regulated gene transcription. *Proc Natl Acad Sci U S A.* 1994;91(18):8517–21. doi: 10.1073/pnas.91.18.8517.
32. Razandi M, Pedram A, Greene GL, Levin ER. Cell membrane and nuclear estrogen receptors (ERs) originate from a single transcript: studies of ER $\alpha$  and ER $\beta$  expressed in Chinese hamster ovary cells. *Mol Endocrinol.* 1999;13(2):307–19. doi: 10.1210/mend.13.2.0239.
33. Lösel R, Wehling M. Nongenomic actions of steroid hormones. *Nat Rev Mol Cell Biol.* 2003;4(1):46–56. doi: 10.1038/nrm1009.
34. Hüser S, Guth S, Joost HG, Soukup ST, Köhrle J, Kreienbrock L, Diel P, Lachenmeier DW, Eisenbrand G, Vollmer G, Nöthlings U, Marko D, Mally A, Grune T, Lehmann L, Steinberg P, Kulling SE. Effects of isoflavones on breast tissue and the thyroid hormone system in humans: a comprehensive safety evaluation. *Arch Toxicol.* 2018;92(9):2703–2748. doi: 10.1007/s00204-018-2279-8.
35. Dixon RA. Phytoestrogens. *Annu Rev Plant Biol.* 2004;55:225–61. doi: 10.1146/annurev.arplant.55.031903.141729.
36. Křížová L, Dadáková K, Kašparovská J, Kašparovský T. Isoflavones. *Molecules.* 2019;24(6):1076. doi: 10.3390/molecules24061076.
37. Kurzer MS, Xu X. Dietary phytoestrogens. *Annu Rev Nutr.* 1997;17:353–81. doi: 10.1146/annurev.nutr.17.1.353.
38. Ionescu VS, Popa A, Alexandru A, Manole E, Neagu M, Pop S. Dietary Phytoestrogens and Their Metabolites as Epigenetic Modulators with Impact on Human Health. *Antioxidants (Basel).* 2021;10(12):1893. doi: 10.3390/antiox10121893.
39. Crozier A, Jaganath IB, Clifford MN. Dietary phenolics: chemistry, bioavailability and effects on health. *Nat Prod Rep.* 2009;26(8):1001–43. doi: 10.1039/b802662a.
40. Kiyama R. Estrogenic flavonoids and their molecular mechanisms of action. *J Nutr Biochem.* 2023;114:109250. doi: 10.1016/j.jnutbio.2022.109250.

41. Vuorelaa P, Leinonenb M, Saikkuc P, Tammela P, Rauhad JP, Wennberge T, Vuorela H. Natural products in the process of finding new drug candidates. *Curr Med Chem.* 2004;11(11):1375–89. doi: 10.2174/0929867043365116.
42. Clifford MN. Diet-derived phenols in plasma and tissues and their implications for health. *Planta Med.* 2004;70(12):1103–14. doi: 10.1055/s-2004-835835.
43. Hüser S, Guth S, Joost HG, Soukup ST, Köhrle J, Kreienbrock L, Diel P, Lachenmeier DW, Eisenbrand G, Vollmer G, Nöthlings U, Marko D, Mally A, Grune T, Lehmann L, Steinberg P, Kulling SE. Effects of isoflavones on breast tissue and the thyroid hormone system in humans: a comprehensive safety evaluation. *Arch Toxicol.* 2018;92(9):2703–2748. doi: 10.1007/s00204-018-2279-8.
44. Cassidy A, Minihane AM. The role of metabolism (and the microbiome) in defining the clinical efficacy of dietary flavonoids. *Am J Clin Nutr.* 2017;105(1):10–22. doi: 10.3945/ajcn.116.136051.
45. Luca SV, Macovei I, Bujor A, Miron A, Skalicka-Woźniak K, Aprotosoae AC, Trifan A. Bioactivity of dietary polyphenols: The role of metabolites. *Crit Rev Food Sci Nutr.* 2020;60(4):626–659. doi: 10.1080/10408398.2018.1546669.
46. Day AJ, DuPont MS, Ridley S, Rhodes M, Rhodes MJ, Morgan MR, Williamson G. Deglycosylation of flavonoid and isoflavonoid glycosides by human small intestine and liver beta-glucosidase activity. *FEBS Lett.* 1998;25;436(1):71–5. doi: 10.1016/s0014-5793(98)01101-6.
47. Setchell KD, Brown NM, Zimmer-Nechemias L, Brashear WT, Wolfe BE, Kirschner AS, Heubi JE. Evidence for lack of absorption of soy isoflavone glycosides in humans, supporting the crucial role of intestinal metabolism for bioavailability. *Am J Clin Nutr.* 2002;76(2):447–53. doi: 10.1093/ajcn/76.2.447.
48. Polkowski K, Mazurek AP. Biological properties of genistein. A review of in vitro and in vivo data. *Acta Pol Pharm.* 2000;57(2):135–55.
49. Kuiper GG, Lemmen JG, Carlsson B, Corton JC, Safe SH, van der Saag PT, van der Burg B, Gustafsson JA. Interaction of estrogenic chemicals and phytoestrogens with estrogen receptor beta. *Endocrinology.* 1998;139(10):4252–63. doi: 10.1210/endo.139.10.6216.
50. Mueller SO, Simon S, Chae K, Metzler M, Korach KS. Phytoestrogens and their human metabolites show distinct agonistic and antagonistic properties on estrogen receptor alpha (ERalpha) and ERbeta in human cells. *Toxicol Sci.* 2004;80(1):14–25. doi: 10.1093/toxsci/kfh147.
51. Nikov GN, Hopkins NE, Boue S, Alworth WL. Interactions of dietary estrogens with human estrogen receptors and the effect on estrogen receptor-estrogen response element complex formation. *Environ Health Perspect.* 2000;108(9):867–72. doi: 10.1289/ehp.00108867.
52. Schrepfer, S.; Deuse, T.; Münzel, T.; Schäfer, H.; Braendle, W.; Reichensperner, H. The selective estrogen receptor-beta agonist biochanin A shows vasculoprotective effects without uterotrophic activity Menopause. 2006;13(3):489–99. doi: 10.1097/01.gme.0000185941.63497.10.
53. Aiyer HS, Warri AM, Woode DR, Hilakivi-Clarke L, Clarke R. Influence of berry polyphenols on receptor signaling and cell-death pathways: implications for breast cancer prevention. *J Agric Food Chem.* 2012;60(23):5693–708. doi: 10.1021/jf204084f.
54. Moutsatsou P. The spectrum of phytoestrogens in nature: our knowledge is expanding. *Hormones (Athens).* 2007;6(3):173–93.
55. van der Woude H, ter Veld MGR, Jacobs N, van der Saag PT, Murk AJ, Rietjens IMCM. The stimulation of cell proliferation by quercetin is mediated by the estrogen receptor. *Mol Nutr Food Res.* 2005;49(8):763–71. doi: 10.1002/mnfr.200500036.
56. Milligan S., Kalita J., Pocock V., Heyerick A., De Cooman L., Rong H., De Keukeleire D. Oestrogenic activity of the hop phyto-oestrogen, 8-prenylnaringenin. *Reprod.-Camb.* 2002;123:235–242. doi: 10.1530/rep.0.1230235.
57. Schaefer O., Hümpel M., Fritzsche K.-H., Bohlmann R., Schleuning W.-D. 8-Prenyl naringenin is a potent ERα selective phytoestrogen present in hops and beer. *J. Steroid Biochem. Mol. Biol.* 2003;84:359–360. doi: 10.1016/S0960-0760(03)00050-5.
58. Penttinen P, Jaehrling J, Damdimopoulos AE, Inzunza J, Lemmen JG, van der Saag P, Pettersson K, Gauglitz G, Makela S, Pongratz I. Diet-derived polyphenol metabolite enterolactone is a tissue-specific estrogen receptor activator *Endocrinology.* 2007;148(10):4875–86. doi: 10.1210/en.2007-0289.
59. Zakic T, Pekovic-Vaughan V, Cvoro A, Korac A, Jankovic A, Korac B. Redox and metabolic reprogramming in breast cancer and cancer-associated adipose tissue. *FEBS Lett.* 2023 Dec 23. doi: 10.1002/1873-3468.14794. Epub ahead of print. PMID: 38140817.
60. Cvoro A, Tzagarakis-Foster C, Tatomer D, Paruthiyil S, Fox MS, Leitman DC. Distinct roles of unliganded and liganded estrogen receptors in transcriptional repression. *Mol Cell.* 2006;21(4):555–64. doi: 10.1016/j.molcel.2006.01.014.
61. An J, Tzagarakis-Foster C, Scharschmidt TC, Lomri N, Leitman DC. Estrogen receptor beta-selective transcriptional activity and recruitment of coregulators by phytoestrogens. *J Biol Chem.* 2001;276(21):17808–14. doi: 10.1074/jbc.M100953200.
62. Li Z, Li J, Mo B, Hu C, Liu H, Qi H, Wang X, Xu J. Genistein induces G2/M cell cycle arrest via stable activation of ERK1/2 pathway in MDA-MB-231 breast cancer cells. *Cell Biol Toxicol.* 2008;24(5):401–9. doi: 10.1007/s10565-008-9054-1.
63. Kładna A, Berczyński P, Kruk I, Piechowska T, Aboul-En-ein HY. Studies on the antioxidant properties of some phytoestrogens. *Luminescence.* 2016;31(6):1201–6. doi: 10.1002/bio.3091.
64. Rüfer CE, Kulling SE. Antioxidant activity of isoflavones and their major metabolites using different in vitro assays. *J Agric Food Chem.* 2006;54(8):2926–31. doi: 10.1021/jf053112o.
65. Borrás C, Gambini J, Gómez-Cabrera MC, Sastre J, Pallardó FV, Mann GE, Viña J. Genistein, a soy isoflavone, up-regulates expression of antioxidant genes: involvement of estrogen receptors, ERK1/2, and NFκappaB. *FASEB J.* 2006;20(12):2136–8. doi: 10.1096/fj.05-5522fj.
66. Prietsch RF, Monte LG, da Silva FA, Beira FT, Del Pino FA, Campos VF, Collares T, Pinto LS, Spanevello RM, Gamaro GD, Braganhol E. Genistein induces apoptosis and autophagy in human breast MCF-7 cells by modulating the expression of proapoptotic factors and oxidative stress enzymes. *Mol Cell Biochem.* 2014;390(1-2):235–42. doi: 10.1007/s11010-014-1974-x.

67. Verdrengh M, Jonsson IM, Holmdahl R, Tarkowski A. Genistein as an anti-inflammatory agent. *Inflamm Res*. 2003;52(8):341–6. doi: 10.1007/s00011-003-1182-8.
68. Bonet-Costa V, Herranz-Pérez V, Blanco-Gandía M, Mas-Bargues C, Inglés M, García-Tarraga P, Rodríguez-Arias M, Miñarro J, Borrás C, García-Verdugo JM, Viña J. Clearing Amyloid- $\beta$  through PPAR $\gamma$ /ApoE Activation by Genistein is a Treatment of Experimental Alzheimer's Disease. *J Alzheimers Dis*. 2016;51(3):701–11. doi: 10.3233/JAD-151020.
69. Leitman DC, Christians U. MF101: a multi-component botanical selective estrogen receptor beta modulator for the treatment of menopausal vasomotor symptoms. *Expert Opin Investig Drugs*. 2012;21(7):1031–42. doi: 10.1517/13543784.2012.685652.
70. Cvoro A, Paruthiyil S, Jones JO, Tzagarakis-Foster C, Clegg NJ, Tatomer D, Medina RT, Tagliaferri M, Schaufele F, Scanlan TS, Diamond MI, Cohen I, Leitman DC. Selective activation of estrogen receptor-beta transcriptional pathways by an herbal extract. *Endocrinology*. 2007;148(2):538–47. doi: 10.1210/en.2006-0803.
71. Paruthiyil S, Cvoro A, Zhao X, Wu Z, Sui Y, Staub RE, Baggett S, Herber CB, Griffin C, Tagliaferri M, Harris HA, Cohen I, Bjeldanes LF, Speed TP, Schaufele F, Leitman DC. Drug and cell type-specific regulation of genes with different classes of estrogen receptor beta-selective agonists. *PLoS One*. 2009;4(7):e6271. doi: 10.1371/journal.pone.0006271.
72. Grady D, Sawaya GF, Johnson KC, Koltun W, Hess R, Vittinghoff E, Kristof M, Tagliaferri M, Cohen I, Ensrud KE. MF101, a selective estrogen receptor beta modulator for the treatment of menopausal hot flashes: a phase II clinical trial. *Menopause*. 2009;16(3):458–65. doi: 10.1097/gme.0b013e31818e64dd.

## Fitoestrogeni kao ligandi estrogenskog receptora

Aleksandra Cvoro

Biološki fakultet, Univerzitet u Beogradu,  
Beograd, Srbija

### Kratak sadržaj

*Estrogenski receptori (ER) su ligandom regulisani transkripcioni faktori koji modulišu esencijalne programe transkripcije aktiviranjem ili reprimiranjem ciljane ekspresije gena. S obzirom na uticaj ER signalizacije na razvoj, metabolizam i fiziologiju, izmenjeno funkcionisanje ER leži u osnovi mno-*

*gih poremećaja, zbog čega su ER odavno prepoznati kao biološke i farmaceutske mete od izuzetne važnosti. Iako je 17 $\beta$ -estradiol (E2) glavni endogeni ER ligand, postoji mnoštvo jedinjenja, uključujući fitoestrogene, koja oponašaju estrogene i takođe aktiviraju ER. Fitoestrogeni su sposobni da se vežu za ER i tako utiču na signalne puteve regulisane estrogenom, izazivajući koliko korisne toliko i štetne posledice. U ovom radu dat je sažet pregled dosadašnjih saznanja iz oblasti fitoestrogena u funkciji alternativnih ER liganada.*

**Ključne reči:** Estrogenski receptor; fitoestrogeni; MF101; genistein.