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Paraquat and other dessicants and bleaching herbicides – their influence on weeds, crops and human and animal health

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SUMMARY

The paper provides an overview of various aspects of desiccants and bleaching herbicides, from different chemical groups, which have in common the drying and bleaching of leaf mass. In weed plants, they can act as total or selective herbicides, and be applied in the mature stages of the crop, where by drying the leaf mass they promote the maturation of some important crops such as sunflower, soybeans, and potatoes. More or less, all these various herbicides are highly or significantly toxic, which greatly complicates their application. In the EU, there was an initiative to ban some of them (e.g. paraquat). However, despite their significant toxicity, we believe they should be considered as an alternative to some other herbicides.

Keywords: desiccants, bipyridyl herbicides, paraquat, weeds, crop plants, toxicity.

INTRODUCTION

Weeds have been encountered by humans since the beginning of the development of agriculture, simply because the selection of cultivated plants is most often carried out by cross-breeding (Quarrie, 1997). This process enhances the quantitative traits of the cultivated

plant, such as crop yield (economically the most important property of cultivated plants), but significantly reduces the competitive ability of cultivated plants against native species that grow around them, which are condidered weeds by definition. Historically, humans have always solved that problem manually, by weeding and other methods of physically removing weeds, so the rapid development of plant physiology and organic chemistry in the first half of the 20th century created the preconditions for a completely different way of solving this important problem. Namely, this knowledge led to the fact that in the years before the Second World War, weeds could be selectively removed chemically, without major negative consequences of those compounds on the health and yield of cultivated plants. After World War II, these promising findings led to systematic research into the synthesis of various (primarily organic) compounds and their testing in terms of weed removal efficiency and selectivity, i.e., the absence of significant negative effects of those phytotoxic compounds on cultivated plants (Corbet et al., 1984; Percival and Baker, 1991). Such an approach soon proved to be promising because while routine application of herbicides (as these phytotoxic compounds are called) leads to a reduction in crop yield due to weed competition of 8-15%, without the use of herbicides the total yield loss can reach up to 50% of the potential crop yield (Percival and Baker, 1991). The introduction into practice of a new agrotechnical measure, chemical crop protection, in addition to its benefits, also led to new unknowns and problems, one of which is the resistance of weeds to previously phytotoxic doses of herbicides, which testifies to the plastic variability of those native plants. This has become a serious problem, as the development of new phytotoxic formulations and herbicidal compounds is very expensive and demanding, while the profit can be small, depending on the economic expediency of using them. Additionally, there is the issue of a rapid development of weed resistance to some classes of herbicides (Percival and Baker, 1991; Travlos et al., 2020). This then entails the need for a more careful study of herbicidal effects, both on plants and on other living organisms and ecosystems, due to health and ecological concerns.

EFFECTS OF PARAQUAT AND OTHER DESICCANTS ON WEEDS

Paraquat (also known as methyl viologen) and its related compound diquat, as bipyridyl molecules (Averina et al., 1991), lead to the desiccation of weeds and partially controlled desiccation of crop leaf mass (for faster ripening), inhibiting the electron transport in Photosystem I (PSI) (Corbet et al., 1984). This is achieved when those electrons, instead of the usual photosynthetic acceptors (FNRS: feredoxine-NADPH oxydo-reduktase; Figure 1), are transferred to oxygen. This transfer occurs through Mehler's reactions or the water-water cycle (Kleczkowski, 1993; Janjić et al., 1994; Asada, 2000) producing reactive oxygen species, that disrupt cellular metabolism, leading to the drying and death of plant foliage.

In addition to the well-known bipyridyl herbicides with desiccation properties, and due to the development of resistance to bipyridyls (Streller et al., 1994; Weaver et al., 2004; Qin et al., 2004; Asaduzzaman et al., 2022; Farago et al., 2022), naturally occurring or due to

genetic modification, mainly due to the increased content of antioxidant small compounds (e.g. polyamines) or antioxidant enzymes (which inactivate reactive oxygen species, thereby preventing phytotoxic processes associated with bipyridyls), a new type of desiccant that induces the Mehler reaction (s.c. water-water cycle) has recently been developed (Gerwiuck et al., 1997).

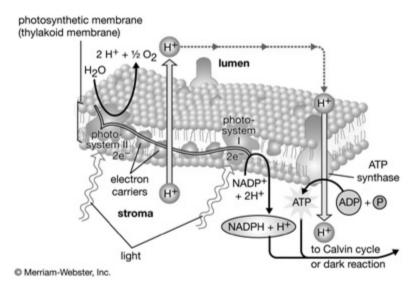


Figure 1. The transport of electrons in the light phase of photosynthesis Slika 1. Transport elektrona u svetloj fazi fotosinteze

In addition to bipyridyls (as PSI inhibitors), which induce Mehler's reaction, there is another type of desiccant herbicides, which at the same time lead to leaf bleaching, indicative of increased reactive oxygen species production. These herbicides primarily function by inhibiting one of the pathways in the biosynthesis of chlorophyll or carotenoids (Sherman et al., 1991; Jacobs et al., 1991; Averina et al., 1991), targeting enzymes such as phytoene desaturase and protoporphyrinogen oxidase. These are diethyl ether, pyridazinone, and furanone type herbicides, as well as amitrole (Corbet et al., 1984; Percival and Baker, 1991). Such an approach, due to the appearance of weeds resistant to bipyridyl, influenced by the emergence of new desiccant herbicides (Ison et al., 2022), such as Voraxor[®], a combination of the herbicides trifludimoxazin and saflufenacil: Tirexor[®] + Kixor[®]. However, it should be mentioned that paraquat, although generally considered a contact herbicide, has been observed to be taken up and transported through plants by the same pathways as polyamines (Fujita and Shinozaki, 2021), which indicates possibly new types of weed resistance to this and other desiccant herbicides (Farago et al., 2022; Lyu et al., 2022). In addition to other treatment practices, the efficacy of paraquat and other desiccants can be enhanced by encapsulation processes (e.g. with chitosan), which prolongs the otherwise short-term contact effect of this herbicide (Kurniadie et al., 2022).

EFFECTS OF PARAQUAT AND OTHER DESICCANTS ON CROP PLANTS

Although the application of herbicides is directed primarily against the harmful effects of weeds on crops, it is essential to also examine their impact on the growth, development, and yield quality and quantity of cultivated plants. This is importand due to the phytotoxic effects of these compounds on crops and possible residues they may leave in the harvested produce, thus entering the food and feed supply as toxic compounds. Therefore, studying the effects of desiccant herbicides is crucial, especially those classified as total herbicides (bipyridyls, amitrol), due to their persistence in both plant products and the soil where they are applied. All in all, it is necessary to distinguish the phytotoxic from the health aspects of the application of desiccant herbicides, when they are used in cultivated plants. The mode of action of desiccant herbicides on these plants varies widely, for example, bipyridyl herbicides such as paraguat, diquat and some related compounds, have many different effects on crop plants. In addition to inducing the production of reactive oxygen species (Asada, 2000), which disrupt cellular metabolism and lead to the decline of crop leaf mass (beneficial in the case of ripening of some crops, such as sunflower, soybean, potato), this group of herbicides causes pheophytinization, i.e. the loss of Mg ions from chlorophyll molecules. This process is another factor in the inhibition of photosynthesis and other metabolic processes and leads to an even greater production of reactive oxygen species (Averina et al., 1991). A series of studies by Nikolić et al. confirmed the pheophytinization of chlorophyll under the effects of diquat (Nikolić and Janjić, 1995; Nikolić et al., 1996a, 1996b; Nikolić, 1997; Pavlović et al., 2014). Additionally, significant changes in photosynthetic membrane proteins were observed under the effects of diguat (Milivojević and Nikolić, 1998, 1999; Milivojević et al., 1997), which were mediated by far-red light (Tables 1 and 2) (Milivojević and Nikolić, 1998). This light is primarily adopted through PSI, the main site of action of bipyridyl herbicides (Ashton and Crafts, 1981).

As indicated in the previous chapter, plant resistance to bipyridyl herbicides can be achieved through enzymatic antioxidant systems or through small molecules with antioxidant properties. This topic was investigated in detail on cultivated plants, and it was found that resistance to bipyridyl herbicides is increased by the action of small molecules (Hart et al., 1993; Donahue et al., 1997; Fujita and Shinozaki, 2021), which probably induce complexation of ions of transitional metals (Fe, Cu, Zn, etc.), as well-known promoters of the Mehler reaction (Chang and Kao, 1997). In addition to the effect on transition metals as mediators of the Mehler reaction, increased resistance to bipyridyl herbicides in cultivated plants is also possible through antioxidant enzyme systems (Kuk et al., 2006; Wang et al., 2021). Since the induction of the production of reactive oxygen species on a small scale has a favorable effect on plant metabolism, it was considered, as in the case of some other types of herbicides (Ashton and Crafts, 1981), that treating crop plants with small doses of bipyridyls might induce the hormesis

Crop/Hoor	Carthean /Cate	Maiza/Kulturuz
intervala (0-48h)	na odnos hlorofila (Chl) i karo	tenoida (Car) u listovima soje i kukuruza
Tabela 1. Efekati	dikvata u mraku ili pod belom	ili daleko-crvenom svetlošću tokom različitih vremenskih
intervals (0-48h) o	on ratios of chlorophylls (Chl) ar	nd carotenoids (Car) in primary leaves of soybean and maize
Table 1. Effects of	liquat in the dark or under "whi	te light" (WL) or far-red radiation (FR) during different time

Crop/Usev			Soybea	n/Soj	a		Maize/Kukuruz											
Parameter Parametar		Chl all	6	C	hl a/C	ar			Chl all	6		Chl a/Car						
Treatment Tretman	0h 24h 48h			0h	24h	48h	5h	12h	18h	36h	48h	5h	12h	18h	36h	48h		
WL	2.1	2.3	2.4	5.3	5.6	4.6	2.6	2.5	2.3	2.5	n.m.	4.9	4.8	4.9	5.5	15.3		
WL+ diquat	2.5	2.0	1.5	4.4	6.6	8.9	2.6	2.6	2.3	1.5	0.8	4.7	5.4	6.1	9.8	35.8		
FR	2.2	2.1	1.7	3.4	3.4	3.9	2.6	2.4	2.4	2.1	2.1	4.9	4.6	4.9	4.6	4.9		
FR+ diquat	2.1	2.1	1.5	3.5	4.4	6.9	2.6	2.4	1.5	1.9	0.9	5.2	4.6	15.8	6.8	11.6		
Dark	2.1	2.3	2.6	5.3	4.1	4.2	2.8	2.7	2.5	1.6	1.2	5.0	4.7	5.2	5.7	6.0		
Dark + diquat	2.3	2.4	2.1	4.1	4.1	5.1	2.7	2.6	2.4	1.4	1.0	4.9	3.6	5.7	13.8	10.7		
LSD o.os		0.07			0.09				0.24					2.13				

n.m.: not measured. According to: Milivojević and Nikolić (1998)

Table 2. Effects of diquat in the dark or under "white light" (WL) or far-red radiation (FR) during different time intervals (0-24h) on polypeptides of photosynthetic reaction centres (RC) and light-harvesting complexes (LHC) of photosystem PSI and PSII of soybean and maize chloroplast thylakoids

Tabela 2. Efekati dikvata u mraku ili pod belom ili daleko-crvenom svetlošću tokom različitih vremenskih intervala (0-48h) na polipeptide fotosintetskih reakcionih centara (RC) i kompleksa za prikupljanje svetlosti (LHC) PSI i FSII tilakoida hloroplasta soje i kukuruza

Crop/Usev	Soybean/Soja											Maize/Kukuruz									
Parameter	PS ₁		PS ₂		LHC		PS ₂ /PS ₁		LHC/PS ₂		PS ₁		PS ₂		LHC		PS ₂ /PS ₁		LHC/PS ₂		
Parametar																					
Treatment	$0h \rightarrow 5h \rightarrow$		>	5 h→		5 h→		$5 h \rightarrow$		$5 h \rightarrow$		5 h→		5 h→		5 h→		5 h→			
Tretman	24	24 h 2		1	24 h		24 h		24 h		24 h		24 h		24 h		24 h		24 h		
WL	7.1	3.0	23.2 3	8.6	35.4	34.3	3.3	1.8	1.7	1.0	6.9	5.2	22.2	33.8	36.7	29.8	3.2	6.5	1.7	0.8	
WL+ diquat	4.0	2.6	19.1 2	25.4	40.7	43.7	4.8	9.8	2.1	1.7	1.9	4.0	24.7	33.6	42.6	25.6	13.0	8.4	1.7	0.8	
FR	5.3	2.6	24.6 2	23.8	23.9	43.7	4.6	9.2	1.0	1.8	2.9	5.8	28.4	25.7	37.1	27.9	9.8	4.4	1.3	1.1	
FR+ diquat	3.5	2.0	12.9 1	5.0	50.7	34.3	3.7	7.5	3.9	1.4	1.2	3.6	21.2	23.4	45.9	23.4	17.7	6.5	2.2	1.0	
Dark	3.5	3.1	19.9 2	22.0	39.0	46.3	5.5	8.7	2.0	1.7	1.5	4.0	25.0	26.3	35.8	30.9	16.7	10.0	1.4	1.2	
Dark + diquat	4.9	1.8	34.9 3	0.0	34.9	37.0	4.7	16.7	1.5	1.2	n.m.	2.6	45.0	32.0	36.3	25.1	45.0	12.3	0.8	0.8	
LSD o.os	0.	0.8 10.5		5	9.3		1.5		1.2		1.1		5.1		8.5		7.6		0.4		

n.m.: not measured. According to: Milivojević and Nikolić (1998)

effect, thereby increasing plant metabolism, which would theoretically have a favorable effect on crop yield. However, these attempts did not give sufficiently promising results (Ferrari et al., 2021), although similar trials have shown favorable results in the model plant of *Arabidopsis thaliana* (Farago et al., 2022). The effects of some other types of desiccant herbicides, lead to leaf bleaching at the same time, indicating an increased production of reactive oxygen species, with mechanisms of action based on the inhibition of one of the pathways in the biosynthesis of chlorophyll or carotenoids. In this case, we can observe the binding of these herbicides to enzymes such as phytoene desaturase and protoporphyrinogen oxidase (Dalla Vekia et al., 2001; di Baccio et al., 2001), which leads to their inhibition, and thus to the increased production of reactive oxygen species. However, there is a physiological difference in the degree of binding of these herbicides to the abovementioned enzymes, which may underline the potential resistance of cultivated plants to their effect (Sherman et al., 1991).

EFFECTS OF PARAQUAT AND OTHER DESICCANTS ON HUMAN AND ANIMAL HEALTH

Paraquat, partly diquat, and other desiccants are highly toxic substances. Paraquat is one of the most poisonous and according to the nomenclature applied in Serbia, classified in the 1st group of poisons (the highest toxicity category). This is partly due to the fact that paraquat induces the production of reactive oxygen species, which disrupt metabolism (Brown and Seither, 1990) and damage human, animal and plant cells. Additionally, paraquat's high volatility can cause eye damage, and diquat can penetrate the skin. Consequently, bipyridyl compounds can easily enter into the body of humans and animals, through means other than inhalation or ingestion of contaminated food or liquids. Desiccant herbicides from other chemical groups mostly cause poisoning by ingestion via contaminated food or liquids, or when they are inadequately applied in the field. Nevertheless, unlike bipyridyls, which are highly toxic, but with a short-term effect due to rapid inactivation, desiccants from other chemical groups are persistent in the environment. This persistence can lead to their accumulation and prolonged toxicity to humans and animals. Detoxification and relief of bipyridyl poisoning symptoms are performed in different ways, either by removing or inactivating transition metal ions (Krall et al., 1991; Sato, 1991), which act as cofactors with bipyridyl in the production of reactive oxygen species, or by using natural (Palipoch et al., 2022; Chen et al., 2023) or synthetic (Li et al., 2021) drugs. The consequences of intoxication with bipyridyl herbicides are severe, causing damage to the eyes, lungs, liver, and kidneys, and potentially leading to long-term symptoms, causing Alzheimer's disease (Marshall and Prior, 2022). That is why, at the first instance of the Court of the European Union, Sweden, with the support of Denmark, Finland and Austria, and against the European Commission, launched an initiative to cancel the license for the use of paraguat as a plant protection product, due to its health risks (Anonymus, 2007). The EU court supported this initiative at that decision-making level.

OTHER ASPECTS RELATING TO PARAQUAT AND OTHER DESICCANTS

Because of these health risks, sophisticated analytical techniques have been developed for the detection of bipyridyl herbicide residues in various human, animal, and plant tissues (Hao et al., 2013; Tsao et al., 2016; Guo et al., 2023). Additionally, various techniques are being considered for the fastest possible inactivation of these highly toxic substances in the event of soil or water contamination (Inthama et al., 2021; Hammami et al., 2022).

CONCLUSIONS AND FUTURE PERSPECTIVES ABOUT THE EFFECTS OF PARAQUAT AND OTHER DESICCANTS IN AGRICULTURE AND RELATED AREAS

Bipyridyl herbicides, along with other desiccants, have shown high efficiency, either for the complete removal of weed vegetation (total herbicides: paraquat, diquat, amitrole), or for its selective removal (e.g. diethyl ether, pyridazinone, and furanone-type herbicides). However, their high toxicity (paraquat) and persistence in the environment increasingly raise concerns, leading to restrictions and even bans on their use. But certainly, despite these restrictions, their high efficiency in removing weeds cannot be ignored. Developed countries, similar to the case of some organochlorine insecticides (e.g. DDT), which are known for their high persistency and a proven harmful effect on the environment, still maintain stocks of these substances for emergency use. This is because having such options available can be crucial for sudden, critical needs. We believe that considering all the limitations in the application of these types of pesticides, our country must also develop a strategy for responding to possible palliative situations, even if it involves the use of these highly toxic herbicides.

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LITERATURE

- Anonymus: Judgment of the Court of First Instance in Case T-229/04. Kingdom of Sweden vs Commission of the European Communities: The court of first instance annuls the directive authorising paraquat as an active plant protection substance. Press release of European Community, 45/07, 2007. https://ec.europa. eu/commission/presscorner/detail/ en/CJE_07_45 and http://curia.europa.eu/jurisp/cgi-bin/form. pl?lang=EN&Submit= recherché&numaff=T-229/04.
- *Asada, K.:* The water-water cycle as alternative photon and electron sinks. Philosophical Transactions of the Royal Society B: Biological Sciences, 355 (1402), 1419-1431, 2000. https://doi:10.1098/rstb.2000.0703.
- Asaduzzaman, M., Koetz, E., Wu, H., Shephard, A.: Paraquat resistance and hormetic response observed in *Conyza sumatrensis* (Retz.). E. Walker (tall feabane) in Australian cotton cropping systems. Phytoparasitica, 50, 269-279, 2022. https://doi.org/10.1007/s1 2600 -021-00956-2.
- Ashton, F. M., Crafts, A. S.: Mode of Action of Herbicides, 2nd Ed. John Wiley & Sons, New York, p. 525, 1981.
- Аверина, Н. Г., Шалыго, N. V., Н. Н.: Линник: Изучение феофотонизации хлорофилла под действием хелаторов металлов пиридинового ряда. Физиология растений, 38 (6), 1059-1065, 1991.
- Brown, O. R., Sejther, R. L.: Paraquat toxicity and pyridine nucleotide coenzyme synthesis: a data correction. Free Radical Biology and Medicined, 8, 113-116, 1990.
- Chang, C. J., Kao, C. H.: Paraquat toxicity is reduced by metal chelators in rice leaves. Physiology Plantarum, 101, 471-476, 1997.

- Chen, H. S., Shen, Y. C., Chi-Cheng Lin, C. C., Juan, C. W., Chang, C. L., Liang, S. Y.: Ethanol-soluble extract of Terminalia chebula attenuates paraquat-induced apoptosis in PC12 cells. Current Topics in Nutraceutical Research, 21 (1), 25-30, 2023. https://doi.org/10.37290/ctnr2641-452X.21:25-30.
- Corbet, J. R., Wright, K., Baillie, A. C.: The Biochemical Mode of Action of Pesticides, Second Edition. Academic Press, London, p. 382, 1984.
- Dalla Vekia, F., Barbato, R., la Rocca, N., Moro, I., Rascio, N.: Responces to bleaching herbicides by leaf chloroplasts of maize plants grown at differnt temperatures. Journal of Experimental Botany, 52 (357), 811-820, 2001.
- di Baccio, D., Quartacci, M. F., Dalla Vekia, F., la Rocca, N., Rascio, N., Navari-Izzo, F.: Bleaching herbicides effects on plastids of dark-grown plants: lipid composition of etioplasts in amitrole and norflurazon-treated barley leaves. Journal of Experimental Botany, 53 (376), 1857-1865, 2001.
- Donahue, J. L., Okpodu, C. M., Cramer, C. L., Grabau, E. A., Alscher, R. G.: Responces of antioxydants to paraquat in pea leaves. Relationship to resistance. Plant Physiology, 113, 249-257, 1997.
- Faragó, D., Zsigmond, L., Benyó, D., Alcazar, R., Rigó, G., Ayaydin, F., Rabilu, S. A., Hunyadi-Gulyás, E., Szabados, L.: Small paraquat resistance proteins modulate paraquat and ABA responses and confer drought tolerance to overexpressing Arabidopsis plants. Plant Cell and Environment, 45, 1985-2003, 2022. https://doi. 10.1111/pce.14338.
- Ferrari, S., de Godoy, D. R. Z., Cunhac, M. L. O., Prado, E. P., Lisboa, L. A. M., dos Santos Cordeiroa, L. F., Carara, L. G. D., de Oliveira, L. C. A.: Can the application of low doses of paraquat induce the hormesis effect in upland rice? Journal of Environmental Science and Health, Part B, 56 (11), 954-961, 2021. https:// doi.org/10. 1080/03601234.2021.1988815.
- Fujita, M., Shinozaki, K.: Identification of polyamine transporters in plants: paraquat transport provides crucial clues. Plant Cell and Physiology, 55 (5), 855-861, 2014. https://doi:10.1093/pcp/pcu032.
- Gerwick, B. C., Fields, S. S., Graupner, P. R., Gray, J. A., Chapin, E. L., Cleveland, J. A., Heim, D. R.: Pyridazocidin, a new microbial phytotoxin with activity in the Mehler reaction. Weed Science, 45, 654-657, 1997.
- Guo, H., Li, L., Gao, L.: Paraquat and diquat: recent updates on their pretreatment and analysis methods since 2010 in biological samples. Molecules, 28, 684, 2023. https://doi.org/10.3390/ molecules28020684.
- Hammami, H., Mozafarjalali, M., Hajiani, M., Nassirli, H.: Removal of paraquat from aqueous solutions by plant extracts as an ecofriendly approach. International Journal of Phytoremediation, 24 (11), 1222-1230, 2022. https://doi.org/10.1080/15226514.2021. 2025037.
- Hao, C., Zhao, X., Morse, D., Yang, P., Taguchi, V., Morra, F.: Optimized liquid chromatography tandem mass spectrometry approach for the determination of diquat and paraquat herbicides. Journal of Chromatography A, 1304, 169-176, 2013. https://doi.org/10.1016/j.chroma.2013.07.033.
- Hart, J. J., DiTomaso, J. M., Kochian, L. V.: Characterization of paraquat transport in protoplasts from maize (Zea mays L.) suspension cells. Plant Physiology, 103, 963-969, 1993.
- Inthama, P., Pumas, P., Pekkoh, J., Pathom-aree, W., Pumas, C.: Plant growth and drought tolerance-promoting bacterium for bioremediation of paraquat pesticide residues in agriculture soils. Frontiers in Microbiology, 12, 604662, 2021. https://doi:10.3389/ fmicb.2021.604662.
- *Ison, R., Francis, I., Readett, G., Brown, M.*: Voraxor[®] Herbicide: An alternative to paraquat in fallow double-knock managing glyphosate and paraquat resistant weeds. In the Proceedings of 2022 Austraasian Weeds Conference, pp. 249-252, 2022.
- Jacobs, J. M., Jacobs, N. J., Sherman, T. D., Duke, S. O.: Effect of Difenil ether herbicides on oxidation of protoporphyrinogen to protoporphyrin in organellar and plasma membrane enriched fractions of barley. Plant Physiology, 97, 197-203, 1991.
- Janjić, V., Veljović-Jovanović, S., Kalezić-Stanković, R., Jovanović, Lj., Marisavljević, D.: Delovanje herbicida na fotosintezu. Acta herbologica, 3 (2), 5-15, 1994.
- *Kleczkowski, L. A.*: Inhibitors of photosynthetic enzymes/carriers and metabolism. Annual Review of Plant Physiology and Plant Molecular Biology, 45, 339-367, 1993.

- Krall, J., Speranza, M. J., Lynch, R. E.: Paraquat-resistant hela cells: increased cellular content of glutathione peroxidase. Archives of Biochemistry and Biophysics, 286 (2), 311-315, 1991.
- Kuk, Y. I., Shin, J. S., Jung, H. L., Guth, J. O., Jung, S., Burgos, N. R.: Mechanism of paraquat tolerance in cucumber leaves of various ages. Weed Science, 54, 6-15, 2006.
- Kurniadie, D., Umiyati, U., Widianto, R., Kato-Noguchi, H.: Effect of chitosan molecules on paraquat herbicidal efficacy under simulated rainfall conditions. Agronomy, 12, 1666, 2022. https://doi.org/10.3390/ agronomy12071666.
- Li, L. R., Chaudhary, B., You, C., Dennis, J. A., Wakeford, H.: Glucocorticoid with cyclophosphamide for oral paraquat poisoning. Cochrane Database of Systematic Reviews. 6 (6), CD008084. 2021. https:// doi:10.1002/14651858. CD008084.pub5.
- Lyu, Y. S., Cao, L. M., Huang, W. Q., Jian-Xiang Liu, J. X., Lu, H. P.: Disruption of three polyamine uptake transporter genes in rice by CRISPR/Cas9 gene editing confers tolerance to herbicide paraquat. aBIOTECH, 3 (2), 140-145, 2022. https://doi.org/10. 1007/s42994-022-00075-4.
- Marshall, C., Prior, M.: UK farmers call for weedkiller ban over Parkinson's fears. BBC News, 2022. https://www.bbc.com/news/science-environment-60836892.
- *Milivojević, D., Nikolić, B.:* Dejstvo herbicida dikvata na pigmente i polipeptide tilakoidnih membrana hloroplasta soje. Zbornik izvoda saopštenja XI-tog Simpozijuma Jugoslovenskog društva za fiziologiju biljaka, Novi Sad, str. 75, 1995.
- Milivojević, D., Nikolić, B.: Effects of herbicide diquat on pigment and polypeptide composistion of thylakoide membranes of soybean chloroplasts. In the Proceedings of 1st Regional Symposium "Chemistry and Environment", Vrnjačka Banja, pp. 643-646, 1996.
- *Milivojević, D., Nikolić, B., Janjić, V.:* The influence of light of different qualities and intensities on the phytotoxic effect of diquat. Acta herbologica, 6 (1), 23-30, 1997.
- *Milivojević, D., Nikolić, B.:* Effects of diquat on pigment-protein complexes of thylakoid membranes in soybean and maize plants. Biology Plantarum, 41 (4), 597-600, 1998.
- *Milivojević, D., Nikolić, B.:* Effects of diquat and metribuzin and different light fluence rates on pigments and PS1 and PS2 Complexes in soybean thylakoid membranes. Archive of Biology Science, Belgrade, 51 (2), 31-32, 1999.
- Nikolić, B., Janjić, V.: Uticaj dikvata na sadržaj fotosintetskih pigmenata lista soje (*Glycine max* Merr.). Zbornik izvoda saopštenja XI-tog Simpozijuma Jugoslovenskog društva za fiziologiju biljaka, str. 74, 1995.
- Nikolić, B., Janić, V., Milivojević, D.: Sinergističko dejstvo svetlosti i nekih herbicida na sadržaj fotosinthtskih pigmenata u listu soje (*Glycine max* Merr.). Acta herbologica, 5 (2), 63-70, 1996a.
- Nikolić, B., Milivojević, D., Janić, V.: The Influence of Some Herbicides on Photosynthetic Pigments of Soybean (*Glycine max* Merr.) Leaf. In the Proceedings of 1st Regional Symposium "Chemistry and Environment", Vrnjačka Banja, pp. 639-642, 1996b.
- Nikolić, B.: Uticaj herbicida metribuzina, linurona i dikvata na sadržaj fotosintetiskih pigmenata listova soje (*Glycine max* Merr.). Magistarski rad, Biološki fakultet, Univerzitet u Beogradu, str. 169, 1997.
- Palipoch, S., Punsawad, C., Koomhin, P., Na-Ek, P., Poonsawat, W., Kimseng, R., Chotipong, P., Bunluepuech, K., Yusakul, G., Suwannalert, P.: Aqueous Thunbergia laurifolia leaf extract alleviates paraquat-induced lung injury in rats by inhibiting oxidative stress and inflammation. BMC Complementary Medicine and Therapies, 22, 83, 2022. https://doi.org/10.1186/s12906-022-03567-4.
- Pavlović, D., Nikolić, B., Đurović, S., Waisi, H., Anđelković, A., Marisavljević, D.: Chlorophyll as a measure of plant health: Agroecological aspects. Pesticides and Phytomedicine, 29 (1), 21-34, 2014. https://doi.https:// doi.org/10.2298/pif.v29i1.5121.
- Percival, M. P., Baker, N. R.: Herbicides and photosynthesis. in: Topics in photosynthesis. Vol 10, Herbicides. Elsevier, Holland, (Baker, N. R. and M. P. Percival, Eds.), pp. 1-21, 1991.
- *Qin, Y., Cairns, A., Powles, S. B.*: Paraquat resistance in a population of *Lolium rigidum*. Functional Plant Biology, 31, 247-254, 2004.

- *Quarrie, S.:* How to use physiology to improve the drought resistance of maize. In the Proceedings of abstracts of 12th Symposium of the Yugoslav Society for Plant Physiology. Kragujevac, Serbia (Ed. Agriculture Research Institute "Serbia"), p. 6, 1997.
- Sato, R., Oshio, H., Koike, H., Inoue, Y., Yoshida, S., Takahashi, N.: Specific Binding of Protoporphyrin IX to a Membrane-Bound 63 Kilodalton Polypeptide in Cucumber Cotyledons Treated withh Diphenyl Ether-Type Herbicides. Plant Physiology, 96, 432-437, 1991.
- Sheryar, Ahmad, S., Chaudhary, B., Azam, M., Ayub, C. M., Yaseen, M.: Comparison of effects of different herbicides on NPK uptake, yield, and quality of Kinnow mandarin plants. Soil and Environment, 41 (1), 75-84, 2022.
- Sherman, T. D., Becerril, J. M., Matsumoto, H., Duke, M. V., Jacobs, J. M., Jacobs, N. J., Duke, S. O.: Physiological basis for differential sensitivies of plant species to protoporphrinogen oxydase-inhibiting herbicides. Plant Physiology, 97, 280-287, 1991.
- Streller, S., Karpinski, S., Halgren, J. E., Wingsle, G.: Four cytosolic-type CuZne-superoxide dismutases in germinating seeds of *Pinus silvestris*. Physiology Plantarum, 92, 443-450, 1994.
- Travlos, I., de Prado, R., Chachalis, D., Bilalis, D. J.: Editorial: Herbicide resistance in weeds: early detection, mechanisms, dispersal, new insights and management issues. Frontiers in Ecology and Evolution, 8, 213, 2020. https://doi.org/10.3389/fevo.2020. 00213.
- Tsao, Y. C., Lai, Y. C., Liu, H. C., Liu, R. H., Lin, D. L: Simultaneous determination and quantitation of paraquat, diquat, glufosinate and glyphosate in postmortem blood and urine by LC–MS-MS. Journal of Analytical Toxicology, 40, 427-436, 2016. https://doi: 10.1093/jat/bkw042.
- Wang, H., Xu, D., Zhu, X., Wang, M., Xia, Z.: The maize SUMO conjugating enzyme ZmSCE1b protects plants from paraquat toxicity. Ecotoxicology and Environmental Safety, 211, 111909, 2021. https://doi.org/10.1016/j. ecoenv.2021.1119 09.
- Weaver, S., Downs, M., Neufeld, B.: Responce of paraquat-resistant and -susceptible horseweed (Conyza canadensis) to diquat, linuron, and oxyfluorfen. Weed Science, 52, 549-553, 2004.

Parakvat i drugi herbicidi – desikanti i izbeljivači – njihov uticaj na korove, useve i zdravlje ljudi i životinja

REZIME

U radu je dat pregled različitih aspekata desikanata i izbeljivačkih herbicida, iz različitih hemijskih grupa, koji imaju zajedničku osobinu isušivanja i izbeljivanja lisne mase. U korovskim biljkama mogu delovati kao totalni ili selektivni herbicidi, a primenjeni u zrelim fazama useva, isušivanjem lisne mase promovišu sazrevanje nekih važnih useva (suncokret, soja, krompir). Većinu ovih herbicida karakteriše visoka ili znatna toksičnost, što znatno komplikuje njihovu primenu. U Evropskoj Uniji došlo je do inicijative za zabranu nekih od ovih aktivnih supstanci (npr. parakvat). Međutim, uprkos njihovoj značajnoj toksičnosti, verujemo da bi ih trebalo razmotriti kao alternativu nekim drugim herbicidima.

Ključne reči: desikanti, bipiridil herbicidi, parakvat, korovi, usevi, toksičnost.