# ROSMARINIC AND CAFFEIC ACID CONTENT AND ANTIOXIDANT POTENTIAL OF THE SALVIA AETHIOPIS L. EXTRACTS

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Aromatic plants are the source of pharmacologically active compounds with high antioxidant effects. Among them, Salvia L. species, sages, have been known world-wide as spices, aromas, medicines, natural preservatives and antioxidant agents since ancient times. Literature data have shown that Salvia aethiopis L. expresses various biological effects. The aim of this research was to determine the quantity of rosmarinic and caffeic acid, usually present in sages, in S. aethiopis extracts prepared with different solvents and to estimate their antioxidant effects. The above-ground parts of S. aethiopis were collected in the period of full blossom in the surrounding area of Niš, Ploče, Serbia. The plant material was air-dried, pulverized and extracted with absolute and 80% methanol, 96%, 80% and 60% ethanol and ethyl acetate (M, M80, E, E80, E60 and EA, respectively) in an ultrasonic bath. The phenolic acids were quantified by High Performance Liquid Chromatography and antioxidant effect was estimated by two complementary in vitro methods: 2,2-diphenyl-1-picrylhydrazyl (DPPH) and  $\beta$ -carotene/linoleic acid (BC) models. Extract E80 contained the highest amount of rosmarinic acid  $(231.09\pm4.11 \mu g/mg)$  and E60 was the richest in caffeic acid (4.39 $\pm$ 0.80  $\mu$ g/mg). M80 was the most efficient in DPPH antioxidant assay, while E60 expressed the best antilipoperoxidant activity in BC method. The presence of significant amount of rosmarinic acid along with caffeic acid and excellent antioxidant activity of the extracts may be contributable to their potential usage in different pathological conditions, especially in the modulation of oxidative stress. Acta Medica Medianae 2017;56(3):121-128.

Key words: Salvia aethiopis L., extracts, rosmarinic acid, caffeic acid, antioxidant

activity

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#### Introduction

The recent upsurge of natural antioxidants usage has raised the importance of aromatic plants due to their safety and benefits that they can offer as extracts, essential oils and spices (1). These plant preparations are the source of pharmacologically active compounds with high antioxidant potentials such as phenolic acids, flavonoids, diterpenes and tannins which can act as reducing agents, hydrogen donators or singlet oxygen quenchers (2, 3). It is well-established that diets rich in the antioxidants have a protective role with anticarcinogenic or cardioprotective effects (3).

Salvia L. species have been known for their medicinal properties and world-wide use as spices, aromas, natural preservatives and antioxidant agents since ancient times (4). Among them, Salvia officinalis L. is most used in traditional medicine and it is attributed with a large number of pharmacological effects such as antioxidant, antiinflammatory, antimicrobial, anticancer, anti-nociceptive, antimutagenic, hypoglycemic and hypolipidemic. This herb is traditionally and mainly used for the treatment of mild dyspepsia, hyperhidrosis, age-related cognitive disorders, and throat and skin inflammations (5). S. officinalis and other sages are rich in polyphenols, especially in phenolic acids and flavonoids (4). The majority of phenolic acids in these species are caffeic acid derivatives. Caffeic acid is the structural basis of different plant metabolites, including simple monomers and a variety of oligomers (6). This phenolic acid manifests a variety of pharmacological effects such as antioxidant, antidepressive, antimetastatic, anti-tumor, hepatoprotective, antiinflammatory, anti-coagulatory, antimicrobial and photoprotective (7-13). The most important compound in sages is rosmarinic acid, a caffeic acid ester, which has been increasingly studied lately in terms of its pharmacological activities (14). Rosmarinic acid has been proven to express antioxidant, anti-inflammatory, antimutagenic, hepatoprotective, antithrombotic, antiplatelet, anti-angiogenic, antibacterial, antiviral (especially against *Herpes simplex* infections), cancer hemopreventive activity, anxiolytic and astringent effects (15-17).

Salvia aethiopis L. is biennial, rarely perennial plant species that occurs sporadically in Serbia inhabiting dry places, meadows and sandy substrates (18). There is not much information on its traditional use except its use as antiflatulent and reconstituent (19). Scientific literature data show that S. aethiopis extracts express antimicrobial, antifungal, antimycobacterial, cytotoxic, antioxidant, anti-cholinesterase and muscarinic receptor activity (20-25). In addition, aethiopinone, an onaphthoquinone diterpenoid, isolated from roots manifested significant antinociceptive, anti-inflammatory and the bleeding time increasing effects (26).

All these facts point to a great therapeutic potential of this plant species, its extracts and isolated compounds. There is a lack of data on rosmarinic and caffeic acid content in S. *aethiopis* extracts, especially from Serbia, which are certainly, in a high percentage, responsible for their activities. Therefore, the aim of this research is directed to the determination of these compounds in S. *aethiopis* extracts prepared with different solvents. Furthermore, the study defines antioxidant effects of the extracts including the determination of the rosmarinic and caffeic acid impact on their manifestation.

### **Material and methods**

### Plant material

The above-ground parts of S. *aethiopis*, Lamiaceae family were collected in the period of full blossom in the wider surrounding area of Niš, Ploče, Serbia. A voucher specimen has been deposited at the Faculty of Science and Mathematics, University of Niš, Department of Biology and Ecology (No. 13198).

### Extracts preparation

The plant material was dried at well-ventilated place protected from direct sunlight and then ground to powder. The powdered material was extracted with absolute and 80% methanol, 96%, 80% and 60% ethanol and ethyl acetate (M, M80, E, E80, E60 and EA, respectively), in 1:10 ratio, in an ultrasonic bath for 40 minutes. Final extracts were obtained after filtration and total removal of the solvents in a rotary vacuum evaporator. The ex-tracts were kept in a sealed bottle in a dark place at 4°C before the experiments.

HPLC (High-performance liquid chromatography) determination of rosmarinic and caffeic acid

HPLC analysis was conducted using a method described by Kostic et al. (2015) (1) with some modifications. The compounds were quantified on Agilent 1200 HPLC system (Agilent Technologies, Palo Alto, Calif., U.S.A.) with a diode array detector (DAD) using the analytical column -Purospher STAR RP-18e (150×4.6 mm) with the particle size of 5 µm. The extracts were dissolved in HPLC methanol to a concentration of 10 mg/mL. The obtained methanolic solutions (10 µL) were injected into the system with column flow of 0.7 mL/min. Operating temperature was adjusted and kept at 30°C. A mobile phase consisted of 0.1% water-trifluoroacetic acid solution (A) and acetonitrile (B) with linear gradient: 0-3 min 5-5% B, 3-32 min 5-28% B, 32-44 min 25-50% B, 44-52 min 50-80% B, 52-54 min 80-90% B, 54-59 min 90-5% B, and 59-60 min 5% B. The quantification of rosmarinic and caffeic acid was performed using calibration curves made with the standards and the final results are expressed as µg of the compound in mg of the extract.

Determination of antioxidant activity

2,2-diphenyl-1-picrylhydrazyl (DPPH) radical scavenging assessment

The antiradical activity of the extracts was evaluated with DPPH method reported by Miladinović et al. (2014) (27). The extracts were initially dissolved in several decreasing concentrations. Forty microliters of the dissolved extract were added to the microtitre plate's well which contained 120  $\mu$ L of ethanol, immediately followed by 40  $\mu$ L of DPPH solution (0.2 mg/mL). The plate was placed in a dark location for 30 minutes after short shaking. The absorbances of the wells' contents were measured using ELISA reader (Multi-skan Ascent No354, Thermo Labsystems, Finland) at 540 nm. The inhibition of free-radical generation (%) was calculated by the equation:

% inhibition =  $[(AK-AA)/(AK-AS)] \times 100$ 

AA is the absorbance of the analyte, AK is the absorbance of the control made up of the solvent and DPPH solution and AS is the absorbance of the solvent. The inhibitory concentration of the extract which neutralized 50% of DPPH radicals ( $IC_{50}$ ) was calculated from a curve constructed with the percentage of inhibition and the concentration of the extract. Ascorbic acid and atocopherol served as positive controls.

 $\beta\mbox{-}Carotene/linoleic acid method for anti-lipoper-oxidation assessment$ 

The ability of the extract to reduce lipid per-oxidation was assessed with the  $\beta\mbox{-}car\mbox{$ 

linoleic acid method described by Miladinovic et al. (2014) (27). Two milligrams of crystalline  $\beta$ carotene was dissolved in 10 mL of chloroform and then 2 mL of this solution were pipetted into a round-bottom flask, followed by 364 µL of Tween 20 and 50 µL of linoleic acid. After the total evaporation of chloroform in a vacuum evaporator, 100 mL of oxigenated water was added with shaking. The resulting emulsion was transferred (200  $\mu$ L) into the wells of microtitre plates which had already contained 25 µL of the extracts dissolved in several decreasing concentrations. A solution without  $\beta$ -carotene was prepared as a blank control. The absorbances of the solution in microtitre plates were immediately measured by ELISA reader (Multiskan Ascent No354, Thermo Labsystems, Finland) at 450 nm after gentle shaking  $(A_0)$ . Microtitre plates were then placed at 55°C in an incubator for 2 hours. After incubation the absorbances were measured again (A120) at the same wavelength. The percentage of lipoperoxidation inhibition was calculated using the equation (28):

% inhibition =  $[(A_{120}/A_0)] \times 100$ 

 $IC_{50}$  of the extract was calculated from a curve which represented the dependence of the concentration and % of inhibition. Ascorbic acid and a-tocopherol served as positive controls.

#### Statistical analysis

The results present mean values of three measurements  $\pm$  standard deviations. The results were analyzed by one-way analysis of variance (ANOVA). Differences among means were compared with Duncan test at p <0.05 considered as significant. Correlations among components and antioxidant activities of the extracts were analyzed following the Pearson's correlation coefficient method (p <0.05). The software package SPSS 17 (SPSS, Inc., Chicago, IL) was used for statistical analyses.

#### Results

The results of the extraction yield and the content of rosmarinic and caffeic acid are shown in Table 1. The best extraction yield was noted for the

**Table 1.** Yields and rosmarinic (RA) and caffeic acid(CA) contents in Salvia aethiopis L. extracts

	yield	RA	CA
extract	%	µg/mg	µg/mg
Е	3.60	140.24±3.21 <sup>a</sup>	1.33±0.03ª
E80	8.45	231.09±4.11 <sup>b</sup>	3.02±0.11 <sup>b</sup>
E60	14.05	72.53±1.05 <sup>c</sup>	4.39±0.80 <sup>c</sup>
М	8.05	222.18±3.01 <sup>d</sup>	1.16±0.05ª
M80	9.20	223.26±2.24 <sup>d</sup>	1.99±0.03 <sup>d</sup>
EA	3.10	5.89±0.33 <sup>e</sup>	$1.00 \pm 0.01^{a}$
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The results present mean values of three measure-ments  $\pm$  standard deviations.

Values in the columns with different lower-case letters are significantly different at p < 0.05.

**Table 2.** Antioxidant potentials of Salvia aethiopis L.extracts estimated by 2,2-diphenyl-1-picrylhydrazyl(DPPH) radical scavenging and  $\beta$ -carotene/linoleic acid(BC) systems

	DDPH	BC
extract	µg/ml	µg/ml
E	32.29±1.83ª	79.54±1.98ª
E80	24.34±0.56 <sup>b</sup>	78.16±0.99ª
E60	47.83±3.58 <sup>c</sup>	37.82±1.45 <sup>b</sup>
М	26.58±0.91 <sup>a,b</sup>	58.86±4.26 <sup>c</sup>
M80	23.79±0.42 <sup>b</sup>	41.66±1.64 <sup>b</sup>
EA	145.64±7.79 <sup>d</sup>	108.29±7.96 <sup>d</sup>
positive controls		
ascorbic acid	4.74±0.34 <sup>e</sup>	22.95±1.52 <sup>9</sup>
a-tocopherol	$10.40 \pm 1.73^{f}$	$0.15 \pm 0.00^{h}$

The results present mean values of three measurements  $\pm$  standard deviations.

Values in the columns with different lower-case letters are significantly different at p < 0.05.

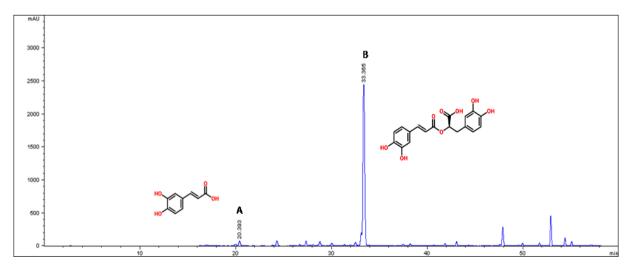


Figure 1. HPLC-chromatogram of the extract E80 of *Salvia aethiopis* L. at 330 nm. Peaks: A – caffeic acid, B – rosmarinic acid

extract E60 made up with the most polar solvent. This extract also contained the highest amount of caffeic acid, while extract E80 contained the greatest quantity of rosmarinic acid (Figure 1). Ethyl acetate was the least efficient in the process of extraction, as well as in the iso-lation of the phenolic acids.

The values of the concentrations of the extracts that inhibited the 50% of free radicals are shown in Table 2. As it is obvious, the extract M80 was the most efficient in DPPH system, immediately followed by E80 and M, while E60 expressed the best antilipoperoxidant activity in  $\beta$ -carotene/linoleic acid method. EA extract was the weakest in both antioxidant methods, as expected. Ascorbic acid and a-tocopherol, positive controls, were uneqaled in free radical reduction.

## Discussion

The polarity of a solvent is the main factor for extraction yield and amount of extracted compounds. However, the extraction technique including temperature, time, pH and chemical structure of the components in the plant material are highly contributable (29). In this study, considering the same extraction technique used for all extracts, the main factors for different yields and quantity of the phenolic acids are their chemical structure, the type of used solvents and change of their polarity. The smallest yield was noted in EA extract due to the lowest solvent's polarity. Concentrated and absolute alcohols gave lower yields compared to their aqueous-alcoholic forms. This implies that adding water, by increasing the polarity, provided better extraction yields and that the plant material contained mostly polar compounds.

The literature data show that S. aethiopis extracts contain rosmarinic, caffeic, p-coumaric, oleanolic, ursolic and chlorogenic acid. Apigenin, apigenin-7-O-glucoside, luteolin, luteolin-7-3',4'trimethylether and salvigenin were identified as the representatives of flavonoids (30-32). Veličković et al. (2002) (22) identified volatile compounds of S. aethiopis extracts, whereby 1,8cineol and a-thujone were most present. It is interesting that there are specific compounds, diterepenes, salvipisone and aethiopinone found in the roots, as well as, sesterterpene lactones, salviaethiopisolide and 13-epi- salviaethiopisolide found in aerial parts of the herb (33-35). Chemical compositions of S. aethiopis essential oils are well studied whereby α-copaen, β-caryophyllene and germacrene D are the most dominant compounds (36).

This study confirmed the presence of rosmarinic and caffeic acid in S. *aethiopis* in our locality, the surroundings of Niš, Serbia. The content of rosmarinic acid in the extracts ranged from 5.89  $\mu$ g/mg to 231.09  $\mu$ g/mg. Ethyl acetate as nonpolar solvent extracted the lowest quantity of rosmarinic acid, as expected, and on the other side, E80 was the richest in its amount. Absolute

and 80% methanols were highly effective solvents for its isolation, as well. It is interesting that the extract E60, prepared with the most polar solvent, contained a low amount of rosmarinic acid compared to other polar solvents. This suggests that rosmarinic acid is less isolated from plant material when highly polar solvent is used too. Compared to the previous researches the contents of rosmarinic acid in our extracts are definitively the highest. Namely, the ethanolic and methanolic leaves extract of Hungarian S. aethiopis contained 1.8 µg/mg and 2.8 µg/mg, respectively (37, 38). Furthermore, methanolic extract of S. aethiopis from Romanian flora contained 1.1189 µg/mg (32). The scope of caffeic acid content in our extracts was 1-4.39 µg/mg. The lowest quantity was determined again in EA extract, but it does not significantly differ from the extract E and M. It is obvious that the addition of water to concentrate ethanol and absolute methanol increased the extraction of caffeic acid, so that its quantity rose in the following order M<E<M80< E80<E60. In the aforementioned extracts of other researchers, the caffeic acid contents were also significantly lower and the values ranged from 0.07-0.1185 µg/mg (32, 37, 38).

The assessment of antioxidant activity was carried out by two complementary methods. DPPH method assessed the ability of the extracts to capture free radicals while  $\beta$ -carotene bleaching model was used to examine their potency to inhibit lipid peroxidation (39). EA extract was the weakest in both model systems, as expected. Extract M80 was the most superior in DPPH system with IC<sub>50</sub> of 23.79 $\pm$ 0.42 µg/ml. On the one hand, E60 was least active in antiradical activity among the extracts prepared with polar solvents. but on the other hand, it had the strongest antilipo-peroxidant activity which is significant from the standpoint of its rosmarinic and caffeic acid content. Statistically significant is the connection between rosmarinic acid and DPPH antioxidant activity (r = -0.853, p < 0.05). This correlation indicates that rosmarinic acid is highly responsible for antiradical activity, in DPPH system, what other researches established as well (1). In contrast, caffeic acid is partially, but statistically significantly, accountable for antilipoperoxidant activity (r =-0.423, p <0.05). Antiradical activity of S. aethiopis extracts investigated by other researchers varied. Firuzi et al. (2013) (21) determined poor activity of the S. aethiopis extract with IC<sub>50</sub> of 237.37  $\mu$ g/ml, while Tepe et al. (2006) (40) even found the total absence of antiradical activity. Best efficacy was performed by the methanolic extract from Turkey which inhibited 68.72% of free radicals with the concentration of 25 µg/ml (24). Great antilipoperoxidant activity was observed in the extract reported by Tosun et al. (2009), whereby 2 g/l of the methanolic extract inhibited 70.40% of linoleic-free radicals (41).

Rosmarinic and caffeic acid have been widely investigated lately in terms of its antioxidant activities. Rosmarinic acid is able to increase the oxidative and physical stability of liposomes (42). In addition, it was found that rosmarinic acid protects a phospholipid membrane against oxidative damage by the reduction of radical propagation (43). The production of reactive oxygen species and release of interleukin (IL)-6 are significantly decreased in the presence of rosmarinic acid, and at the same time it can prevent UVB damage of human keratinocytes (44). The antioxidant effect of rosmarinic acid is considered to be involved in the prevention of DNA damage induced by doxorubicin. It is important that it reduces the frequency of micronuclei and does not express genotoxic effects (45). Osakabe et al. (2002) (46) suggests that rosmarinic acid could act as a liver protector due to the scavenging and reducing activities of superoxide or peroxynitirite in mice hepatocytes. Due to the ability to scavenge peroxynitrites, rosmarinic acid may protect against cognitive dysfunction in mice induced by amyloid beta protein which is in direct connection with Alzhemier disease (47). In addition to its neuroprotective effect, it is shown that rosmarinic acid significantly suppresses H<sub>2</sub>O<sub>2</sub>-induced reactive oxygen species (ROS) formation in human dopaminergic cell line, SH-SY5Y, and rats astrocytes (48, 49). Its antioxidant activity might be responsible for anti-angiogenic potential due to the reduction of ROS-associated expression of vascular endothelial growth factor and release of IL-8 in vitro (16). The treatment with rosmarinic acid diminishes pancreatic β-cell dysfunction and glucolipotoxicity-mediated oxidative stress in high-fat diet streptozotocin (STZ)-induced type 2 diabetes in albino rats possibly thanks to its antioxidant effect (50). It is also responsible for the alleviation of the ROS production in human gingival fibroblasts pretreated with lipopolysaccharide (51). Caffeic acid is also an effective antioxidant agent in biochemical reactions. It significantly de-creases the effects of pro-oxidants such as iron, sodium nitroprusside and quinolinic acid which causes increase in the malondialdehyde contents of the

brain (52, 53). Caffeic acid inhibits the oxidative stress induced by H<sub>2</sub>O<sub>2</sub> and suppresses the IL-8 secretion and its mRNA expression in human intestinal epithelial Caco-2 cells (54). Khan et al. (2012) (55) concluded that caffeic acid attenuated 12-O-tetradecanoyl-phorbol-13-acetate-induced lipid peroxidation in mouse skin along with the expression of nuclear factor kappa B and cyclooxygenase-2 and tumor necrosis factor-a release suggesting its possible anticarcinogenic activity. It has a protective role in ethanol metabolism-induced oxidative damage in SK-Hep-1 cells by blocking ROS production and enhancing antioxidant potentials (56). Caffeic acid may be beneficial in the attenuation of iron nitrilotriacetate- and nickel-induced oxidative damage in the rat kidney and liver, respectively (57, 58). Nardini et al. (1998) demonstrated that this phenolic acid is able to reduce glutathione depletion and lipid peroxidation in U937 human monocytic cells exposed to t-butyl hydroperoxide (59).

## Conclusion

Considering the reported results, S. aethiopis extracts are the important sources of, primarily, rosmarinic acid along with caffeic acid. The presence of these phenolics in the diet may play an important role in the modulation of oxidative reactions. An extract with particularly high level of rosmarinic acid is E80, while extract E60 contains the largest amount of caffeic acid. All extracts, except ethyl acetate, were superior in antioxidant activities expressions in both models. Therefore, S. aethiopis extracts, made up with polar solutions, could be of great importance for usage in the conditions where oxidative reactions have signi-ficant destabilizing and pathological role. Further investigations are recommended for in vivo anti-oxidant testing of the extracts, and for the identifi-cation and quantification of other present com-pounds as well.

#### References

- Kostić M, Zlatković B, Miladinović B, Živanović S, Mihajilov-Krstev T, Pavlović D, et al. Rosmarinic acid levels, phenolic contents, antioxidant and antimicrobial activities of the extracts from *Salvia verbenaca* L. obtained with different solvents and procedures. J Food Biochem 2015; 39(2):199-208. [CrossRef]
- Erkan N, Ayranci G, Ayranci E. Antioxidant activities of rosemary (*Rosmarinus officinalis* L.) extract, blackseed (*Nigella sativa* L.) essential oil, carnosic acid, rosmarinic acid and sesamol. Food Chem 2008; 110(1):76-82. [CrossRef] [PubMed]
- Rice-Evans CA, Miller NJ, Paganga G. Structureantioxidant activity relationships of flavonoids and phenolic acids. Free Radical Bio Medicine 1996; 20(7):933-56. [CrossRef] [PubMed]
- Deans GS, Simpson JME. Antioxidants from *Salvia* officinalis. In: Kintzios SE, editor. Sage the genus *Salvia*. Amsterdam: Harwood Academic Publishers; 2000. p. 185-92.
- Ghorbani A, Esmaeilizadeh M. Pharmacological properties of *Salvia officinalis* and its components. J Tradit Complement Med. In press 2017. [CrossRef]
- Kamatou GP, Viljoen AM, Steenkamp P. Antioxidant, antiinflammatory activities and HPLC analysis of South African Salvia species. Food Chem 2010; 119(2):684-8. [CrossRef]
- Gülçin İ. Antioxidant activity of caffeic acid (3, 4dihydroxycinnamic acid). Toxicology 2006; 217(2):213-20. [CrossRef] [PubMed]
- Takeda H, Tsuji M, Inazu M, Egashira T, Matsumiya T. Rosmarinic acid and caffeic acid produce antidepressive-like effect in the forced swimming test in mice. Eur J Pharmacol 2002; 449(3):261-7. [CrossRef] [PubMed]
- Chung TW, Moon SK, Chang YC, Ko JH, Lee YC, Cho G, et al. Novel and therapeutic effect of caffeic acid and caffeic acid phenyl ester on hepatocarcinoma cells: complete regression of hepatoma growth and metastasis by dual mechanism. FASEB J 2004; 18(14):1670-81. [CrossRef] [PubMed]
- Janbaz KH, Saeed SA, Gilani AH. Studies on the protective effects of caffeic acid and quercetin on chemical-induced hepatotoxicity in rodents. Phyto medicine 2004; 11(5):424-30. [CrossRef] [PubMed]
- 11. Chao PC, Hsu CC, Yin MC. Anti-inflammatory and anti-coagulatory activities of caffeic acid and ellagic acid in cardiac tissue of diabetic mice. Nutr metab (Lond) 2009; 6(1):33. [CrossRef] [PubMed]
- Aziz NH, Farag SE, Mousa LA, Abo-Zaid MA. Comparative antibacterial and antifungal effects of some phenolic compounds. Microbios 1997; 93(374):43-54. [PubMed]
- Saija A, Tomaino A, Trombetta D, De Pasquale A, Uccella N, Barbuzzi T, et al. *In vitro* and *in vivo* evaluation of caffeic and ferulic acids as topical photoprotective agents. Int J Pharm 2000; 199(1):39-47. [CrossRef] [PubMed]
- 14. Bulgakov VP, Inyushkina YV, Fedoreyev SA. Rosmarinic acid and its derivatives: biotechnology and applications. Crit Rev Biotechnol 2012; 32(3):203-17. [CrossRef] [PubMed]

- 15. Zou ZW, Xu LN, Tian JY. Antithrombotic and antiplatelet effects of rosmarinic acid, a watersoluble component isolated from radix *Salviae miltiorrhizae* (danshen). Yao Xue Xue Bao 1992; 28(4):241-5. [PubMed]
- 16. Huang SS, Zheng RL. Rosmarinic acid inhibits angiogenesis and its mechanism of action *in vitro*. Cancer letters 2006; 239(2): 271-80. [CrossRef] [PubMed]
- 17. Petersen M, Simmonds MSJ. Rosmarinic acid. Phytochemistry 2003; 62(2):121–5. [CrossRef] [PubMed]
- Lima CF, Fernandes-Ferreira M, Pereira-Wilson C. Phenolic compounds protect HepG2 cells from oxidative damage: Relevance of glutathione levels. Life Sci 2006; 79(21):2056–68. [CrossRef] [PubMed]
- 19. Diklic N. Rod *Salvia* L. In: Josifović M, editor. Flora SR Srbije, 6. Beograd: Srpska Akademija Nauka i Umetnosti; 1974. p. 444-5.
- Naghibi F, Mosaddegh M, Mohammadi Motamed M, Ghorbani A. Labiatae family in folk medicine in Iran: from ethnobotany to pharmacology. Iran J Pharm Res 2010; 4(2):63-79.
- Firuzi O, Miri R, Asadollahi M, Eslami S, Jassbi AR. Cytotoxic, antioxidant and antimicrobial activities and phenolic contents of eleven *Salvia* species from Iran. Iran J Pharm Res 2013; 12(4):801-10. [PubMed]
- Veličković DT, Ranđelović NV, Ristić MS, Šmelcerović AA, Veličković AS. Chemical composition and antimicrobial action of the ethanol extracts of *Salvia pratensis* L. *Salvia glutinosa* L. and *Salvia aethiopis* L. J Serb Chem Soc 2002; 67(10):639-46.
- Tosun F, Kizilay CA, Sener B, Vural M, Palittapongarnpim P. Antimycobacterial screening of some Turkish plants. J Ethnopharmacol 2004: 95(2-3):273-5. [CrossRef] [PubMed]
- 24. Senol FS, Orhan I, Celep F, Kahraman A, Doğan M, Yilmaz G, et al. Survey of 55 Turkish *Salvia* taxa for their acetylcholinesterase inhibitory and antioxidant activities. Food Chem 2010; 120(1):34-43. [CrossRef]
- Wake G, Court J, Pickering A, Lewis R, Wilkins R, Perry E. CNS acetylcholine receptor activity in European medicinal plants traditionally used to improve failing memory. J Ethnopharmacol 2000; 69(2):105-14. [CrossRef] [PubMed]
- 26. Hernández-Pérez M, Rabanal RM, de la Torre MC, Rodríguez B. Analgesic, anti-inflammatory, antipyretic and haematological effects of aethiopinone, an o-naphthoquinone diterpenoid from Salvia aethiopis roots and two hemisynthetic derivatives. Planta Med 1995; 61(6):505-9. [CrossRef] [PubMed]
- 27. Miladinović B, Kostić M, Šavikin K, Đorđević B, Mihajilov-Krstev T, Živanović S, et al. Chemical profile and antioxidative and antimicrobial activity of juices and extracts of 4 black currants varieties (*Ribes nigrum* L.). J Food Sci 2014; 79(3):C301-9. [CrossRef] [PubMed]
- 28. Barros L, Ferreira MJ, Queiros B, Ferreira IC, Baptista P. Total phenols, ascorbic acid,  $\beta$ -carotene

and lycopene in Portuguese wild edible mushrooms and their antioxidant activities. Food Chem 2007; 103(2):413-9. [CrossRef]

- 29. López A, Rico M, Rivero A, de Tangil MS. The effects of solvents on the phenolic contents and antioxidant activity of *Stypocaulon scoparium* algae extracts. Food Chem 2011; 125(3):1104-9. [CrossRef]
- 30. Lu Y, Foo LY. Polyphenolics of *Salvia*—a review. Phytochemistry 2002; 59(2):117-40. [<u>CrossRef</u>] [<u>PubMed</u>]
- Janicsák G, Veres K, Kakasy AZ, Máthé I. Study of the oleanolic and ursolic acid contents of some species of the Lamiaceae. Biochem System Ecol 2006; 34(5):392-6. [CrossRef]
- 32. Coisin M, Necula R, Grigoras V, Gille E, Rosenhech E, Zamfirache MM. Phytochemical evaluation of some *Salvia* species from Romanian flora. Analele Stiintifice ale Universitatii" Analele Stiintifice ale Universitatii "Al. I. Cuza" din Iasi 2012; 58(1):35-44.
- 33. Rodríguez B, Fernández-Gadea F, Savona G. A rearranged abietane diterpenoid from the root of *Salvia aethiopis*. Phytochemistry 1984; 23(8):1805-6. [<u>CrossRef</u>]
- 34. Gonzalez MS, San Segundo JM, Grande MC, Medarde M, Bellido IS. Sesterterpene lactones from *Salvia aethiopis*. Salviaethiopisolide and 13-episalviaethiopisolide. Tetrahedron 1989; 45(11): 3575-82. [CrossRef]
- 35. Boya MT, Valverde S. An orthoquinone isolated from *Salvia aethiopis*. Phytochemistry 1981; 20(6):1367-8. [CrossRef]
- 36. Coisin M, Burzo I, Stefan M, Rosenhech E, Zamfirache MM. Chemical composition and antibacterial activity of essential oils of three *Salvia* species, widespread in Eastern Romania. Analele Stiintifice ale Universitatii "Al. I. Cuza" din Iasi 2012; 58(1):51-8.
- Janicsák G, Máthé I, Miklóssy-Vári V, Blunden G. Comparative studies of the rosmarinic and caffeic acid contents of Lamiaceae species. Biochem System Ecol 1999; 27(7):733-8. [CrossRef]
- 38. Zupkó I, Hohmann J, Rédei D, Falkay G, Janicsák G, Máthé I. Antioxidant activity of leaves of *Salvia* species in enzyme-dependent and enzyme-independent systems of lipid peroxidation and their phenolic constituents. Planta Med 2001; 67(4):366-8. [CrossRef] [PubMed]
- 39. Koleva II, van Beek TA, Linssen JP, Groot AD, Evstatieva LN. Screening of plant extracts for antioxidant activity: a comparative study on three testing methods. Phytochem Anal 2002; 13(1):8-17. [CrossRef] [PubMed]
- 40. Tepe B, Sokmen M, Akpulat HA, Sokmen A. Screening of the antioxidant potentials of six *Salvia* species from Turkey. Food Chem 2006; 95(2):200-4. [CrossRef]
- 41. Tosun M, Ercisli S, Sengul M, Ozer H, Polat T, Ozturk E. Antioxidant properties and total phenolic content of eight *Salvia* species from Turkey. Biol Res 2009; 42(2):175-81. [CrossRef] [PubMed]
- 42. Panya A, Laguerre M, Lecomte J, Villeneuve P, Weiss J, McClements DJ, et al. Effects of chitosan and rosmarinate esters on the physical and oxidative stability of liposomes. J Agr Food Chem 2010; 58(9):5679-84. [CrossRef] [PubMed]
- 43. Pérez-Fons L , GarzÓn MT , Micol V. Relationship between the antioxidant capacity and effect of rosemary (*Rosmarinus officinalis* L.) polyphenols on membrane phospholipid order. J Agr Food Chem 2009; 58(1):161-71. [CrossRef] [PubMed]

- 44. Vostálová J, Zdařilová A, Svobodová A. Prunella vulgaris extract and rosmarinic acid prevent UVBinduced DNA damage and oxidative stress in HaCaT keratinocytes. Arch Dermatol Res 2010; 302(3):171-81. [CrossRef] [PubMed]
- 45. Furtado RA, De Araujo FR, Resende FA, Cunha WR, Tavares DC. Protective effect of rosmarinic acid on V79 cells evaluated by the micronucleus and comet assays. J Appl Toxicolo 2010; 30(3):254-9. [PubMed]
- 46. Osakabe N, Yasuda A, Natsume M, Sanbongi C, Kato Y, Osawa T, et al. Rosmarinic acid, a major polyphenolic component of *Perilla frutescens*, reduces lipopolysaccharide (LPS)-induced liver injury in D-galactosamine (D-GalN)-sensitized mice. Free Radical Bio Med 2002; 33(6):798-806. [CrossRef] [PubMed]
- 47. Alkam T, Nitta A, Mizoguchi H, Itoh A, Nabeshima T. A natural scavenger of peroxynitrites, rosmarinic acid, protects against impairment of memory induced by Aβ 25–35. Behav Brain Res 2007; 180(2):139-45. [CrossRef] [PubMed]
- 48. Lee HJ, Cho HS, Park E, Kim S, Lee SY, Kim CS, et al. Rosmarinic acid protects human dopaminergic neuronal cells against hydrogen peroxide-induced apoptosis. Toxicology 2008; 250(2-3):109-15. [CrossRef] [PubMed]
- 49. Gao LP, Wei HL, Zhao HS, Xiao SY, Zheng RL. Antiapoptotic and antioxidant effects of rosmarinic acid in astrocytes. Pharmazie 2005; 60(1):62-5. [PubMed]
- 50. Govindaraj J, Sorimuthu Pillai S. Rosmarinic acid modulates the antioxidant status and protects pancreatic tissues from glucolipotoxicity mediated oxidative stress in high-fat diet: streptozotocininduced diabetic rats. Mol Cell biochem 2015; 404(1-2):143-59. [CrossRef] [PubMed]
- 51. Zdařilová A, Svobodová A, Šimánek V, Ulrichová J. Prunella vulgaris extract and rosmarinic acid suppress lipopolysaccharide-induced alteration in human gingival fibroblasts. Toxicol in Vitro 2009; 23(3):386-92. [CrossRef] [PubMed]
- 52. Oboh G, Agunloye OM, Akinyemi AJ, Ademiluyi AO, Adefegha SA. Comparative study on the inhibitory effect of caffeic and chlorogenic acids on key enzymes linked to Alzheimer's disease and some pro-oxidant induced oxidative stress in rats' brain*in vitro*. Neurochem Res 2013; 38(2):413-9. [CrossRef] [PubMed]
- 53. Lafay S, Gueux E, Rayssiguier Y, Mazur A, Rémésy C, Scalbert A. Caffeic acid inhibits oxidative stress and reduces hypercholesterolemia induced by iron overload in rats. Int J Vitam Nutr Res 2005; 75(2):119-25. [CrossRef] [PubMed]
- 54. Zhao Z, Shin HS, Satsu H, Totsuka M, Shimizu M. 5-Caffeoylquinic acid and caffeic acid down-regulate the oxidative stress-and TNF-*a*-induced secretion of interleukin-8 from Caco-2 cells. J Agr Food Chem 2008; 56(10):3863-8. [CrossRef] [PubMed]
- 55. Khan AQ, Khan R, Qamar W, Lateef A, Ali F, Tahir M, et al. Caffeic acid attenuates 12-Otetradecanoyl-phorbol-13-acetate (TPA)-induced NF-κB and COX-2 expression in mouse skin: abrogation of oxidative stress, inflammatory responses and proinflammatory cytokine production. Food Chem Toxicol 2012; 50(2):175-83. [CrossRef] [PubMed]
- 56. Lee KM, Kang HS, Yun CH, Kwak HS. Potential in vitro protective effect of quercetin, catechin, caffeic acid and phytic acid against ethanol-induced oxidative stress in SK-Hep-1 cells. Biomol Ther (Seoul) 2012; 20(5):492-8. [CrossRef] [PubMed]

- 57. Rehman MU, Sultana S. Attenuation of oxidative stress, inflammation and early markers of tumor promotion by caffeic acid in Fe-NTA exposed kidneys of Wistar rats. Mol Cell Biochem 2011; 357(1-2):115. [CrossRef] [PubMed]
- 58. Pari L, Prasath A. Efficacy of caffeic acid in preventing nickel induced oxidative damage in liver

of rats. Chem-Biol Interact 2008; 173(2):77-83. [CrossRef] [PubMed]

59. Nardini M, Pisu P, Gentili V, Natella F, Di M, Piccolella F, et al. Effect of caffeic acid on tert-butyl hydroperoxide-induced oxidative stress in U937. Free Radical Bio Med 1998; 25(9):1098-105. [CrossRef] [PubMed]

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# SADRŽAJ RUZMARINSKE I KAFENE KISELINE I ANTIOKSIDATIVNI POTENCIJAL EKSTRAKTA BILJNE VRSTE SALVIA AETHIOPIS L.

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Aromatične biljne vrste predstavljaju izvor farmakološki aktivnih jedinjenja sa visokim antioksidativni delovanjem. Među njima su Salvia L. vrste, žalfije, od davnina poznate širom sveta kao začini, arome, lekovita sredstva, prirodni konzervansi i antioksidansi. Podaci iz literature pokazuju da Salvia aethiopis L. ispoljava različite biološke efekte. Cilj ovog istraživanja bio je utvrditi količinu ruzmarinske i kafene kiseline, često zastupljene u žalfijama, u ekstraktima biljne vrste S. aethiopis, pripremljeni sa različitim rastvaračima i proceniti njihove antioksidativne efekte. Nadzemni delovi S. aethiopis su prikupljeni u periodu cvetanja u okolini Niša, Ploče, Srbija. Biljni materijal je osušen na vazduhu, pulverizovan i ekstrahovan pomoću apsolutnog i 80% metanola, 96%, 80% i 60% etanola i etilacetata (M, M80, E, E80, E60 i EA, redom) u ultrazvučnom kupatilu. Fenolne kiseline su kvantifikovane tečnom hromatografijom visokih performansi, dok je antioksidativni efekat procenjen pomoću dve in vitro komplementarne metode: 2,2-difenil-1-pikrilhidrazil (DPPH) i β-karoten/linolne kiseline (BC). Ekstrakt E80 sadržao je najveću količinu ruzmarinske kiseline (231,09±4,11 µg/mg), a E60 je bio najbogatiji u količini kafene kiseline (4,39±0,80 µg/mg). M80 je bio najefikasniji u DPPH testu, dok je E60 izrazio najbolju antilipoperoksidnu aktivnost u BC modelu. Prisustvo značajnih količina ruzmarinske kiseline, uz kafenu kiselinu, i odlična antioksidativna aktivnost, mogu značajno doprineti potencijalnoj upotrebi ekstrakata u različitim patološkim stanjima, posebno u modulaciji oksidativnog stresa. Acta Medica Medianae 2017;56(3):121-128.

**Ključne reči:** Salvia aethiopis L., ekstrakti, ruzmarinska kiselina, kafena kiselina, antioksidativna aktivnost