ISSN 1452-8258

J Med Biochem 42: 249-257, 2023

Original paper Originalni naučni rad

SERUM FETUIN-A AND RANKL LEVELS IN PATIENTS WITH EARLY STAGE BREAST CANCER

SERUM FETUIN-A I NIVOI RANKLA KOD PACIJENATA SA RANIM STADIJOM RAKA DOJKE

Cigdem Usul Afsar¹, Hale Aral², Orçun Can¹, Didem Can Trabulus³, Didem Karacetin⁴, Mehmet Ali Nazlı⁵, Rıza Umar Gursu⁶, Senem Karabulut⁷

 ¹Istinye University Medical Faculty, Liv Vadi Istanbul Hospital, Department of Internal Medicine and Medical Oncology, Istanbul, Turkey
 ²University of Health Sciences, Istanbul Training and Research Hospital, Department of Medical Biochemistry, Istanbul, Turkey
 ³Goztepe Medical Park Hospital, Department of General Surgery, Istanbul, Turkey
 ⁴University of Health Sciences, Cam and Sakura Education and Research Hospital, Department of Radiation Oncology, Istanbul, Turkey
 ⁵University of Health Sciences, Cam and Sakura Education and Research Hospital, Department of Radiation Oncology, Istanbul, Turkey
 ⁶Acıbadem Bakirkoy Hospital, Department of Medical Oncology, Istanbul, Turkey

Summary

Background: Breast cancer (BC) is the primary cause of mortality due to cancer in females around the world. Fetuin-A is known to increase metastases over signals and peroxisomes related with growing. Receptor activator of nuclear factor- κ B ligand (RANKL) takes part in cell adhesion, and RANKL inhibition is used in the management of cancer. We aimed to examine the relationship between serum fetuin-A, RANKL levels, other laboratory parameters and clinical findings in women diagnosed with early stage BC, in our population.

Methods: Women having early stage BC (n=117) met our study inclusion criteria as they had no any anti-cancer therapy before. Thirty-seven healthy women controls were also confirmed with breast examination and ultrasonography and/or mammography according to their ages. Serum samples were stored at -80 °C and analysed via ELISA.

Results: Median age of the patients was 53 (range: 57–86) while it was 47 (range: 23–74) in the healthy group.

Uvod: Rak dojke (BC) je primarni uzrok smrtnosti od raka kod žena širom sveta. Poznato je da fetuin-A povećava metastaze u odnosu na signale i peroksizome povezane sa rastom. Receptorski aktivator liganda nuklearnog faktora-ĸB (RANKL) učestvuje u ćelijskoj adheziji, a RANKL inhibicija se koristi u lečenju kancera. Cilj nam je bio da ispitamo odnos između serumskog fetuina-A, nivoa RANKL, drugih laboratorijskih parametara i kliničkih nalaza kod žena sa dijagno-zom ranog stadijuma BC, u našoj populaciji.

Metode: Žene sa ranim stadijumom BC (n=117) ispunjavale su kriterijume za uključivanje u našu studiju pošto ranije nisu imale nikakvu terapiju protiv raka. Kontrola 37 zdravih žena je takođe potvrđena pregledom dojki i ultrasonografijom i/ili mamografijom u skladu sa njihovim godinama. Uzorci seruma su čuvani na -80 °C i analizirani pomoću ELISA.

Rezultati: Srednja starost pacijenata bila je 53 godine (raspon: 57–86) dok je u zdravoj grupi bila 47 (raspon: 23–

Orçun Can, Asist, Prof

Istinye University Medical Faculty, Department of Internal Medicine and Medical Oncology, Teyyareci Sami Sk. No.3, 34010, Zeytinburnu, Istanbul, Turkey e-mail: drorcuncan@gmail.com

Kratak sadržaj

Address for correspondence:

All authors contributed substantially to the planning, drafting and final revision of the article

Patients had lower high-density lipoprotein levels (p=0.002) and higher neutrophil counts (p=0.014). Fetuin-A and RANKL levels did not differ between the groups (p=0.116 and p=0.439, respectively) but RANKL leves were found to be lower in the favorable histological subtypes (p=0.04).

Conclusions: In this study, we found no correlation between serum fetuin-A levels and clinical findings in patients diagnosed with early stage BC. However, RANKL levels are found to be lower in subgroups with favorable histopathologic subtypes such as tubular, papillary and mucinous BC and there was statistically significant difference.

Keywords: breast cancer, fetuin-A, RANKL, serum

Introduction

Breast cancer (BC) is the second most frequently diagnosed malignancy just behind lung cancer and also the primary cause of mortality due to cancer in female around the world. Over 1.5 million women (25% of all women with cancer) are diagnosed with BC every year throughout the world (1).Today, the total number of BC patients have increased in response to exposure to several risk factors such as abnormal levels of estrogen, smoking, alcohol and obesity (2). Nevertheless, we know that early BC detection could reduce BC death rates significantly in the longterm (3).

Therefore, specific screening methods with the use of the correct biomarkers are important in the detection of early stage BC (4). Specially, blood, saliva, and urine were considered as ideal origin in which to assign the presence of cancer biomarkers such as annexin, peroxiredoxin and calreticulin (5).

Fetuin-A, also called Alpha 2-Heremans Schmid Glycoprotein (AHSG), is a serum glycoprotein synthesized by the liver and secreted into the blood stream (6). Its founded principal role is the inhibition of ectopic calcification, but mounting evidence suggests that it is a multifunctional protein capable of modulating a number of critical signaling pathways and it has roles in disease processes such as diabetes mellitus and kidney disease (7). In particular, high fetuin-A concentrations are found to be associated with atherogenic lipid profile and metabolic syndrome, low fetuin-A levels are related to vascular calcifications and inflammation (8). In addition, there are few suggesting increased fetuin-A level may be a new serum biomarker in early BC (9).

Receptor activator of nuclear factor-kappa B ligand (RANKL) appertain to tumour necrosis factor superfamily which is a group of proteins that act as bidirectional signalling molecules. RANKL with osteocyte origin induces bone destruction by stimulating osteoclasts, while RANKL released from osteoblasts functions in the reverse effect (10). At the same time, available evidence suggests that the RANKL signaling system is associated with in almost all steps in BC development, from primary oncogenesis to the establishment of secondary tumors in the bone (10). 74). Pacijenti su imali niži nivo lipoproteina visoke gustine (p=0,002) i veći broj neutrofila (p=0,014). Nivoi fetuina-A i RANKL nisu se razlikovali između grupa (p=0,116 i p=0,439, respektivno), ali je utvrđeno da su nivoi RANKL niži kod povoljnih histoloških podtipova (p=0,04).

Zaključak: U ovoj studiji nismo pronašli korelaciju između nivoa fetuina-A u serumu i kliničkih nalaza kod pacijenata sa dijagnozom ranog stadijuma BC. Međutim, utvrđeno je da su nivoi RANKL niži u podgrupama sa povoljnim histopatološkim podtipovima kao što su tubularni, papilarni i mucinozni BC i postojala je statistički značajna razlika.

Ključne reči: rak dojke, fetuin-A, RANKL, serum

The potential role of tumor markers is to improve early cancer determination and they significantly provide an unmatched opportunity to understand the disease's biology, improve diagnosis and enhance treatment. In the present study; we aimed to investigate the the serum levels of fetuin-A and RANKL and their relationship with clinical parameters in patients with early stage BC.

Materials and Methods

Demographic features of patients

A total of 117 female patients with early stage BC between the ages of 27-86 years (median: 53) were included in the study. Our control group was 37 healthy women between the ages of 23-74 (median: 47) years. BC was diagnosed according to the ultrasonography and histopathologic findings of the patients. Healthy controls were also confirmed with breast examination and ultrasonography and/or mammography according to their ages. The patients were asked for their medical history (type 2 diabetes, hypertension, smoking and medications) and measured for their glomerular filtration rate (GFR) and other biochemical parameters. Exclusion criteria included trauma history, major surgical history, chronic kidney disease with a creatinine clearance under 15 mL/min, hepatitis (alcoholic, toxic hepatitis, chronic autoimmune), fatty liver, alcoholic and primer biliary cirrhosis, chronic inflammatory disease, acute infection, known lung or liver disease, known rheumatic heart valve disease, congenital heart disease including bicuspid aorta, dilated cardiomyopathy and known osteoporosis.

Serum preparation

Blood was drawn after 12 hours of fasting in the morning. Serum was obtained after at least 30 minutes of clotting by centrifugation at 2500xg for 15 minutes. Serum was stored at -80 °C until assayed. All icteric or haemolytic blood samples were discarded. All parameters were analyzed in all samples together in a single batch at the termination of the experimental protocol (control and patient samples were analysed in the same batch).

Measurement of serum fetuin-A and RANKL levels

Serum fetuin-A and RANKL levels of patients with BC and of the control group were measured in the venous blood. A commercial kit (Assaypro, USA, cat no: EG 63501-1), based on a quantitative sandwich ELISA, was used and results were determined with ELX 800 UV version ELISA reader and calculated in grams per liter. Mean intra-assay and interassay coefficients of variation were less than 4.9% (n:10) and 6.7% (n:10).

Measurement of other biochemical parameters

Levels of serum glucose, urea, creatinine, total cholesterol, HDL, LDL, AST, ALT, GGT, LDH, ALP, total protein, albumin, parathyroid hormone (PTH), calcium, phosphorus, magnesium, CRP, complete blood count (CBC), CEA, CA 15.3 and erythrocyte sedimentation rate (ESR) were measured in the patient and control groups using the same biochemistry laboratory in our hospital.

Statistical analysis

Statistical analyses (Mann–Whitney U-test, Student t test) were performed with SPSS 19 (Statistical Package for Social Sciences). The difference in various parameters were analyzed by the Chi-square test. Pearson correlation test was used for correlating fetuin-A, RANKL and the different biochemical parameters. Multivariate logistic regression model was performed to determine the effect of independent risk factors for BC. P-values < 0.05 were considered significant.

Results

Median age of the patients was 53 (range: 57-86) while it was 47 (range: 23-74) in the healthy group. Patients were 56% postmenopausal, 40% premenopausal and 4% perimenopausal. Twenty-four (20.5%) of the patients had cerbb2 3 positive or cerb2 2 positive and SISH/FISH positive disease. Grade 2 disease was found in 60 (52.6%) patients and 47 patients (41.2%) had grade 3 disease. Seventy-four (63.2%) patients had invasive ductal carcinoma (IDC), 15 (12.8%) had invasive lobular carcinoma (ILC), 6 (5.1%) had mixed (IDC+ILC) carcinoma, 1 had metaplastic cancer and 21 (17.9%) had other favorable (tubular, apocrine, papillary and mucinous) types. Twenty-nine patients (24.7%) had stage I, thirty (25.6%) had stage II and fifty-eight (49.5%) patients had stage III BC.

There was no statistically significant difference between ER status, PR status, cerbb2 status, grade, lymphovascular invasion, perineural invasion, stage, menopausal status and serum parameters. Patients had lower high-density lipoprotein levels (p=0.002) and higher neutrophil counts (p=0.014) rather than the control group (*IA* and *IB*). Fetuin-A and RANKL levels did not differ between the patients and control groups (p=0.116 and p=0.439, respectively) (*Table II*). There was no statistically significant difference in fetuin-A levels according to various clinical/laboratory

Table IA Laboratory values, cell adhesion markers and age of the patients' and control group.

		N	Mean	Std. Dev.	Min	Max	р	units
A.c.o.	Control	37	47.65	11.106	23	74	014	
Age	BC	117	53.21	12.026	27	86	1.014	
	Total	154	51.88	12.015	23	86		
	Control	37	.35137	.096804	.215	.674	116	g/L
Fetuin-A (g/L)	BC	117	.38076	.099044	.224	.743	1 .110	
	Total	154	.37370	.099000	.215	.743		
RANKL (pM)	Control	37	382.895	326.0093	82.9	1763.7	120	рМ
	BC	117	447.340	470.5775	61.1 2486.6		.435	
	Total	154	431.856	440.0707	61.1	2486.6		
	Control	37	5.8492	0.7293	4.2185	35 7.7154		mmol/L
Glucose (mmol/L)	BC	117	6.2067	2.1565	3.8854	21.7584	.524	
	Total	154	6.1207	1.9169	3.8854	21.7584		
	Control	37	23.08	9.8388	8.1585	49.1175	247	mmol/L
Urea (mmol/L)	BC	117	26.1372	14.9833	6.4935	83.4166	.247	
	Total	154	25.4029	13.9537	6.4935	83.4166		
	Control	37	55.83445	11.2394	39.7808	84.8656	279	mmol/L
Creatinine (mmol/L)	BC	117	58.354	12.5857	34.4767	101.662	.2/5	
	Total	154	57.744	12.287	34.4767	101.662		

	Control	37	5.4354	0.9677	3.3364	8.1469	0.81/	mmol/L
Total cholesterol (mmol/L)	BC	117	5.3865	1.1391	3.1553	8.7418	0.014	
	Total	154	5.3984	1.0975	3.1553	8.7418		
	Control	37	1.475	0.341	0.7242	2.276	0.002	mmol/L
HDL-Chol (mmol/L)	BC	117	1.2784	0.3279	0.6466	2.1208	0.002	
	Total	154	1.3255	0.3416	0.6466	2.276		
	Control	37	3.2435	0.7428	1.7587	5.0071	0.654	mmol/L
LDL-Chol (mmol/L)	BC	117	3.3232	0.9931	0.6052	6.6727	- 0.054	
	Total	154	3.3041	0.9374	0.6052	6.6727		
	Control	37	21.78	6.872	14	54	0.260	U/L
AST (U/L)	BC	117	20.58	5.339	11	43	0.200	
	Total	154	20.87	5.744	11	54		
	Control	37	21.24	15.673	9	87	0.260	U/L
ALT (U/L)	BC	117	19.02	8.484	5	56	- 0.269	
	Total	154	19.55	10.643	5	87		
	Control	37	21.03	14.544	8	84	0.407	U/L
GGT (U/L)	BC	117	23.53	16.180	6	104	- 0.405	
	Total	154	22.93	15.793	6	104		
LDH (U/L)	Control	37	178.59	33.815	80	263	0 777	U/L
	BC	117	180.79	42.218	20	296	- 0.775	
	Total	154	180.27	40.265	20	296		
	Control	36	78.08	23.285	40	143	0 5 5 7	U/L
ALP (U/L)	BC	116	80.79	24.051	40	205	- 0.555	
	Total	152	80.15	23.823	40	205		
	Control	37	74.435	6.4162	68.0	107.0	0.050	g/L
Total protein (g/L)	BC	116	72.614	72.614 4.5792 60.7 81.		81.6	- 0.059	
	Total	153	73.054	5.1213	60.7	107.0		
	Control	37	44.089	2.2737	37.5	48.1	0.463	g/L
Albumin (g/L)	BC	117	43.708	2.8838	35.8	52.3	0.405	
	Total	154	43.799	2.7474	35.8	52.3		
	Control	37	2.3851	0.0822	2.2456	2.5201	0 201	mmol/L
Calcium (mmol/L)	BC	117	2.3582	0.1191	1.8963	2.8195	0.201	
	Total	154	2.3646	0.1117	1.8963	2.8195		
	Control	37	1.0448	0.1542	0.7129	1.3419	997	mmol/L
Phosphorus (mmol/L)	BC	116	1.0445	0.1788	0.6161	1.5839		
	Total	153	1.0446	0.1726	0.6161	1.5839		
	Control	37	0.7791	0.0669	0.0669 0.6213 0		0 222	mmol/L
Magnesium (mmol/L)	BC	116	0.7951	0.0718	0.5842	0.9545	0.202	
	Total	153	0.7912	0.0707	0.5842	0.9545		

Table	IB Laboratory	values, ce	ell adhesion	markers	and age o	of the patients'	and control group	э.

	Control	37	58.981	24.8303	22.8	121.9	0.571	ng/L
PTH (ng/L)	BC	117	62.193	31.3949	11.9	213.5	0.571	
	Total	154	61.421	29.9040	11.9	213.5		
	Control	37	22.84	14.299	1	72	0.91/	mm/h
Sedimentation (mm/h)	BC	110	23.11	12.883	4	76	0.914	
	Total	147	23.04	13.204	1	76		
	Control	37	36.7514	36.1676	4.9524	148.9524	0 2 1 0	nmol/L
CRP (nmol/L)	BC	117	47.3524	61.0316	2.0952	434.2857	0.519	
	Total	154	44.7924	56.1467	2.0952	434.2857		
	Control	37	6.7614	1.10867	4.43	9.45	0.096	10 ⁹ /L
WBC (10 ⁹ /L)	BC	117	7.3162	1.84842	3.63	13.12	0.000	
	Total	154	7.1829	1.71353	3.63	13.12		
	Control	37	3.9443	1.00572	1.90	5.82	0.014	10 ⁹ /L
Neutrophil (10 ⁹ /L)	BC	117	4.6630	1.66258	2.00	10.53	0.014	
	Total	154	4.4903	1.55839	1.90	10.53		
	Control	37	2.1519	.50326	1.25	3.41	0 3 2 8	10 ⁹ /L
Lymphocyte (10 ⁹ /L)	BC	117	2.0429	.61292	1.05	4.41	0.520	
	Total	154	2.0691	.58873	1.05	4.41		
	Control	37	264.32	49.311	160	356	0.805	10 ⁹ /L
PLT (10 ⁹ /L)	BC	117	265.88	65.808	123	524	0.895	
	Total	154	265.51	62.097	123	524		
	Control	37	10.408	.8958	8.7	13.8	0.708	FI
MPV (FI)	BC	115	10.481	1.0618	8.4	14.2	0.708	
	Total	152	10.463	1.0215	8.4	14.2		
	Control	5	15.940	5.6145	10.7	24.1	0.777	U/mL
Ca 15.3 (U/mL)	BC	104	22.993	45.8327	3.0	392.6	0.755	
	Total	109	22.670	44.7968	3.0	392.6		
	Control	4	3.117	4.6920	.3	10.1	0.854	mg/L
CEA (μg/L)	BC	103	3.864	8.0352	.3	56.4	0.054	
(~g/ =/	Total	107	3.836	7.9228	.3	56.4		

 Table IC
 Laboratory values, cell adhesion markers and age of the patients' and control group.

Table II Serum fetuin-A and RANKL levels in the patients' and the control group.

	Ν	Mean	Standard deviation	Minimum	Maximum	р
Fetuin-A (g/L)	Control 37 BC 117 Total 154	0.35137 0.38076 0.37370	0.096804 0.099044 0.99000	0.215 0.224 0.215	0.674 0.743 0.743	0.116
RANKL (pM)	Control 37 BC 117 Total 154	Control 37382.895BC 117447.340Total 154431.856		82.9 61.1 61.1	1763.7 2486.6 2486.6	0.439

Variables	Fetuin Median (Range)	n	Р
Ki-67			
≤ 30	0.345 (0.235–0.522)	43 71	0.09
> 30	0.389 (0.224–0.743)	71	
ER			
Negative	0.398 (0.264–0.692)	25	0.38
Positive	0.375 (0.224–0.743)	92	
PR			
Negative	0.399 (0.247–0.692)	30	0.38
Positive	0.373 (0.224–0.743)	87	
HER2			
Negative	0.362 (0.224–0.743)	90	0.71
Positive	0.369 (0.249–0.692)	24	
Classification			
Luminal	0.358 (0.224–0.743)	92	0.39
Others	0.392 (0.264–0.692) 9225	25	
Classification			
Triple positive	0.345 (0.249–0.497)	13	0.40
Others	0.368 (0.224–0.743)	100	
Classification			
Triple negative	0.336 (0.267–0.591)	11	0.63
Others	0.362 (0.224–0.743)	106	
Classification			
HER2 enriched	0.369 (0.249–0.692)	24	0.71
Others	0.362 (0.224–0.743)	90	
Histological subgroups			
Favorable groups*	0.355 (0.242–0.526) 0.355 (0.242-0.526)	24	0.51
Unfavorable groups**	0.361 (0.224–0.743)	93	
Stage			
Early stage	0.360 (0.224–0.650)	60	0.55
Local advanced stage	0.370 (0.235–0.743)	54	

Table III Comparisons of fetuin-A levels according to various clinical/laboratory param
--

Significant p values (less than 0.05) are highlighted in bold.* Apocrine, papillary, mucinous, metaplastic, neuroendocrine, tubular. ** Invasive ductal carcinoma, invasive lobular carcinoma, mixed type.

Variables	RANKL Median (Range)	n	Р
Ki-67			
≤ 30	203.300 (61.100–1577.500)	43	0.16
> 30	273.300 (101.800–2486.600)	71	
ER			
Negative	490.888 (101.800–2486.600)	25	0.54
Positive	434.228 (61.100–1890.500)	92	
PR			
Negative	441.337 (101.800–2486.600)	30	0.66
Positive	448.059 (61.100–1890.500)	87	
HER2			
Negative	240.950 (61.100–1973.600)	90	0.61
Positive	276.600 (101.800–2486.600)	24	
Classification			
Luminal	250.250 (61.100–1890.500)	92	0.54
Others	294.500 (101.800–2486.600)	25	
Classification			
Triple positive	314.600 (121.700–956.700)	13100	0.22
Others	239.250 (61.100–2486.600)		
Classification			
Triple negative	372.900 (113.200–1973.600)	11	0.50
Others	250.250 (61.100–2486.600)	106	
Classification			
HER2 enriched	276.600 (101.800–2486.600)	24	0.61
Others	240.950 (61.100–1973.600)	90	
Histological subgroups			
Favorable groups*	184.500 (80.500–638.300)	24	0.04
Unfavorable groups**	269.100 (61.100-2486.600)	93	
Stage			
I and II	254.600 (61.100–1973.600)	60	0.62
	261.450 (101.800–2486.600)	54	

Table	e IV	Comparisons	of	R/	٩N	ΚL	levels	accordi	ng to	various	clinical	/lat	ooratory	[,] parameters
-------	------	-------------	----	----	----	----	--------	---------	-------	---------	----------	------	----------	-------------------------

Significant p values (less than 0.05) are highlighted in bold. * Apocrine, papillary, mucinous, metaplastic, neuroendocrine, tubular. ** Invasive ductal carcinoma, invasive lobular carcinoma, mixed type.

parameters (*Table III*). However, patients with favorable histopathologies such as tubular, apocrine, papillary and mucinous subtypes (n=24) had lower RANKL values and it was found to be significant (p=0.04) (*Table IV*).

Discussion

For BC, different serum markers were evaluated upto now and some of them are found to be prognostic, some are diagnostic and/or predictive (11). Fetuin-A was originally discovered to be an inhibitor of vascular calcification. Furthermore it is demonstrated that it plays an important role in free fatty acid induced insulin resistance in the liver (12, 13). Increased fetuin-A had been also been linked to increased occurrence of non-alcoholic fatty liver disease and cardiovascular events, believed to be due to its proinflammatory effects. Thus, in contrast it has some anti-inflammatory properties. It is a negative acute-phase reactant in sepsis, promotes wound healing, and is neuroprotective (14). The potential role of fetuin-A in tumor progression stemmed from earlier studies that suggested that it was the cell attachment factor in serum (15). In head and neck squamous cell carcinoma (HNSCC), there was an increased expression of a higher molecular weight fetuin-A (16). There is ectopic synthesis of fetuin-A by divergent cancer cell lines (17). Patients with high ectopic expression of fetuin-A in lung cancer and gastric cancer tend to have lower survival (18, 19). Fetuin-A is an important marker in the tumor microenvironment, for cancer stem cells and for matrix metalloproteinases (20, 21).

Fetuin-A is found to be a serum biomarker for colorectal cancer patients (22). It is found to be increased in malignant pleural effusion of lung cancer patients (23). Furthermore, in a study done in Mexican BC population, the presence of serum autoantibodies against fetuin-A protein found to be useful as serum biomarkers for early-stage BC screening (24). Fetuin-A seems to be a serum chemo-attractant protein that also promotes invasion of BC tumor cells (25).

In our study, we found no association of serum fetuin-A levels for BC patients with other laboratory parameters and with control subjects. This may be a result of exploring only early staged patients. In the Mexican BC population (24), there were 36 patients (30 with ductal and 6 lobular carcinoma) but they used an immune proteomic approach, combining two-dimensional (2D) electrophoresis, Western blot, and matrix-associated laser desorption/ionizationmass spectrometry (MALDI-MS) methods. We used one method which was the ELISA method for the detection of fetuin-A levels. We performed this study in 117 patients with invasive ductal, lobular, tubular, papillary and mucinous cancers. However, in our study, there was a trend to be lower for fetuin-A levels for more favorable histologic subtypes. It is very well known that BC has different histologic subtypes as well as its diffrenet molecular characteristics. In the

References

- Coleman MP, Quaresma M, Berrino F, Lutz JM, Angelis RD, Capocaccia R, et al. Cancer survival in five continents: a worldwide population-based study (CON-CORD). Lancet Oncol 2008; 9(8): 730–56.
- Sun YS, Zhao Z, Yang ZN, Xu F, Lu HJ, Zhu ZY, et al. Risk Factors and Preventions of Breast Cancer. Int J Biol Sci 2017; 13(11): 1387–97.
- Migowski A. Early detection of breast cancer and the interpretation of results of survival studies. Cienc Saude Coletiva 2015; 20: 1309.
- Clinical practice guidelines for the use of tumor markers in breast and colorectal cancer. Adopted on May 17, 1996 by the American Society of Clinical Oncology. J Clin Oncol 1996; 14: 2843–77.

Mexican study (24), there is no data about the tumors' molecular characteristics such as ER, PR and cerbb2 status. In our study; there are older patients than the other study. Taken together all these discrepancies, in our study which was done in Turkish BC patients, fetuin-A levels did not differ.

RANKL/RANK system is seen as a downstream mediator of progesterone-driven mammary epithelial cells proliferation, BC initiation and progression. Expression of RANKL, RANK has been detected in BC cell lines and in human primary BCs. To date, dysregulation of RANKL/RANK at the skeletal level has been widely documented in the context of metastatic bone disease (26). The interference with the RANK/RANKL system could therefore serve as a potential target for prevention and treatment of BC (27, 28). For metastatic BC patients, specifically for patients with bone metastasis, RANKL levels were found to be diagnostic and somewhat predictive for therapy (27). In our study; for early staged BC patients, RANKL levels were found to be lower in the favorable histological subtypes of BC. This is a new topic for early stage BC patients.

Conclusion

We found a correlation between serum RANKL levels and favorable histological subtypes of BC. However, there was no significance between fetuin-A levels and other clinical/laboratory parameters. Further and detailed studies can enlighten the role of these cell adhesion markers better for BC patients.

Conflict of interest statement

All the authors declare that they have no conflict of interest in this work.

- Yi JK, Chang JW, Han W, Lee JW, Ko E, Kim DH, et al. Cancer Epidemiol Biomarkers Prev 2009: 18; 1357–64.
- Dabrowska AM, Tarach JS, Wojtysiak-Duma B, Duma D. Fetuin-A (AHSG) and its usefulness in clinical practice. Review of the literature. Biomed Pap Med Fac Univ Palacky 2015: 159; 352–9.
- 7. Ochieng J, Nangami G, Sakwe A, Moye C, Alvarez J, Whalen D, et al. Impact of Fetuin-A (AHSG) on Tumor Progression and Type 2 Diabetes. Int J Mol Sci 2018; 19: 2211.
- Ix JH, Shlipak MG, Brandenburg VM, Ali S, Ketteler M, Whooley MA. Association between human fetuin-A and the metabolic syndrome: data from the Heart and Soul Study. Circulation 2006; 113: 1760–7.

- Fernández-Grijalva AL, Aguilar-Lemarroy A, Jave-Suarez LF, Gutiérrez-Ortega A, Godinez-Melgoza PA, Herrera-Rodríguez SE, et al. Alpha 2HS-glycoprotein, a tumorassociated antigen (TAA) detected in Mexican patients with early-stage breast cancer. J Proteomics 2015; 112: 301–12.
- Ono T, Hayashi M, Sasaki F, Nakashima T. RANKL biology: bone metabolism, the immune system, and beyond. Inflamm Regen 2020; 40: 2.
- Aglan SA, Elsammak M, Elsammak O, El-Bakoury EA, Elsheredy HG, Ahmed YS, Sultan MH, Awad AM. Evaluation of serum Nestin and HOTAIR rs12826786 C>T polymorphism as screening tools for breast cancer in Egyptian women. J Med Biochem 2021; 40 (1): 17– 25.
- Trepanowski JF, J Mey J, Varady KA. Fetuin-A: A Novel Link Between Obesity and Related Complications. Int J Obes (Lond) 2015; 39(5): 734–41.
- Ochieng J, Korolkova OY, Li G, Jin R, Chen Z, Matusik RJ, et al. Fetuin-A Promotes 3-Dimensional Growth in LNCaP Prostate Cancer Cells by Sequestering Extracellular Vesicles to Their Surfaces to Act as Signaling Platforms. Int J Mol Sci 2022; 23(7): 4031.
- Mukhopadhyay S, Mondal SA, Kumar M, Dutta D. Proinflammatory and antiinflammatory attributes of fetuin-a: a novel hepatokine modulating cardiovascular and glycemic outcomes in metabolic syndrome. Endocr Pract 2014; 20(12): 1345–51.
- Fisher HW, Puck TT, Sato G. Molecular growth requirements of single mammalian cells: The action of fetuin in promoting cell attachment to glass. Proc Natl Acad Sci 1958; 44: 4–10.
- Arnaud P, Miribel L, Emerson DL. α2-HS glycoprotein. Methods Enzymol 1988; 163: 431–41.
- Mintz PJ, Rietz AC, Cardo-Vila M, Ozawa MG, Dondossola E, Do KA, et al. Discovery and horizontal follow-up of an autoantibody signature in human prostate cancer. Proc Natl Acad Sci 2015; 112: 2515–20.
- Gyorffy B, Surowiak P, Budczies J, Lanczky A. Online survival analysis software to assess the prognostic value of biomarkers using transcriptomic data in non-small-cell lung cancer. PLoS ONE 2013; 8: e82241.
- Szasz AM, Lanczky A, Nagy A, Forster S, Hark K, Green JE, et al. Cross-validation of survival associated biomark-

ers in gastric cancer using transcriptomic data of 1,065 patients. Oncotarget 2016; 7: 49322–33.

- Ochieng J, Nangami G, Sakwe A, Moye C, Alvarez J, Whalen D, et al. Impact of Fetuin-A (AHSG) on Tumor Progression and Type 2 Diabetes. Int J Mol Sci 2018; 19(8): E2211.
- Dong Y, Ding D, Gu J, Chen M, Li S. Alpha-2 Heremans Schmid Glycoprotein (AHSG) promotes the proliferation of bladder cancer cells by regulating the TGF-β signalling pathway. Bioengineered 2022; 13(6): 14282–98.
- 22. Fan NJ, Kang R, Ge XY, Li M, Liu Y, Chen HM, et al. Identification α2-HS glycoprotein precursor and tubulin β-chain as serology diagnosis biomarker of colorectal cancer. Diagn Pathol 2014; 9: 53.
- Yu CJ, Wang CL, Wang CI, Chen CD, Dan YM, Wu CC, et al. Comprehensive proteome analysis of malignant pleural effusion for lung cancer biomarker discovery by using multidimensional protein identification technology. J Proteome Res 2011; 10: 4671–82.
- Fernández-Grijalva AL, Aguilar-Lemarroy A, Jave-Suarez LF, Gutiérrez-Ortega A, Godinez-Melgoza PA, Herrera-Rodríguez SE, et al. Alpha 2HS-glycoprotein, a tumorassociated antigen (TAA) detected in Mexican patients with early-stage breast cancer. J Proteomics 2015; 112: 301–12.
- 25. Nangami GN, Watson K, Parker-Johnson K, Okereke KO, Sakwe A, Thompson P, et al. Fetuin-A (α2HS-glyco-protein) is a serum chemo-attractant that also promotes invasion of tumor cells through Matrigel. Biochem Biophys Res Commun 2013; 438(4): 660–5.
- Infante M, Fabi A, Cognetti F, Gorini S, Caprio M, Fabbri A. RANKL/RANK/OPG system beyond bone remodeling: involvement in breast cancer and clinical perspectives. J Exp Clin Cancer Res 2019; 38(1): 12.
- 27. Kiesel L, Kohl A. Role of the RANK/RANKL pathway in breast cancer. Maturitas 2016; 86: 10–6.
- Mularczyk M, Bourebaba Y, Kowalczuk A, Marycz K, Bourebaba L. Probiotics-rich emulsion improves insulin signalling in Palmitate/Oleate-challenged human hepatocarcinoma cells through the modulation of Fetuin-A/TLR4-JNK-NF-κB pathway. Biomed Pharmacother 2021; 139: 111560.

Received: April 15, 2022 Accepted: October 01, 2022