THE DEVELOPMENT AND VALIDATION OF HPLC METHOD FOR QUANTIFICATION OF DL- α -TOCOPHEROL IN QUINOA SEEDS (*Chenopodium quinoa* Willd.)

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ABSTRACT: Aiming to achieve a precise method with easy application, this study presents the development and validation of a fast method for quantification of vitamin DL- α -tocopherol in quinoa seeds. The methodology was based on an extraction procedure using ultrasonic bath and determination by normal-phase HPLC with UV-VIS detector. Validation parameters showed adequate linearity, relative standard deviations between 0.5 and 0.8% (n=10), limits of detection and quantification were 3.0 ng/mL and 11.0 ng/mL, respectively. Moreover, testing the robustness of the method suggested that it was not changeable with time or condition. The results showed that this method is accurate and simple and thus applicable in laboratories for determination of DL- α -tocopherol in quinoa seeds.

Key words: DL- α -tocopherol, quinoa seed, HPLC

INTRODUCTION

In recent years, there has been a great interest in the relationship between plant-based nutrition and the prevention of human diseases caused by increased levels of radicals (Frias et al., 2005).

Vitamin E is known as an essential micro-nutrient for maintaining the health and wellbeing of humans due to its antioxidant activities (Cadenas and Packer, 2002; Brigelius-Flohe and Galli, 2010). DL- α -tocopherol, the form of vitamin E that is preferentially absorbed and accumulated in humans, is a fat-soluble antioxidant that acts as a peroxyl radical scavengers preventing the propagation of free radicals in tissues. DL- α -tocopherol reacts with free radicals thus forming a tocopheryl radical, which is subsequently reduced by a hydrogen donor and therefore returned to its reduced form (Traber and Stevens, 2011).

As it is fat-soluble, mentioned vitamin is incorporated into cell membranes, has a protective role in terms of oxidative damage and thus plays an important role in prevention of Alzheimer's disease and cancer (Tucker and Townsend, 2005).

While cereal grains are known to be good source of vitamin E, scientists strive towards finding alternative sources of vitamin E that can promote health. The pseudocereal quinoa (*Chenopodium quinoa* Willd.), which is native to the Andean regions of South America, has good nutritional profile and is considered as health-promoting food (Matiacevich et al., 2006). High nutritional value of quinoa seeds is related to the high protein content and wide range of minerals and vitamins (Fleming and Galwey, 1995). Koziol (1992) reported that quinoa contains more

α -tocopherol than rice, barley and wheat (approximately 5.37 mg/100 g of DM).

Quinoa flour of excellent quality is obtained from quinoa seeds and nowadays widely used as a substitution for wheat flour, for instance, in bread, noodles, pasta and sweet biscuits (Valencia-Chamorro, 2003). Quinoa flour can also be extruded, ensuring food with physical, sensorial and nutritional qualities at an enviable level (Valencia-Chamorro, 2003).

The aim of this study was to develop and validate a rapid and accurate method for quantification of DL- α -tocopherol in quinoa seeds. Such study would contribute to further knowledge relating to the stability of this vitamin in different products obtained from quinoa seeds.

MATERIALS AND METHODS

Quinoa seeds

Ten genotypes of quinoa seeds (*Chenopodium quinoa* Willd.) were obtained from field experiment conducted in province Vojvodina (north Serbia).

Extraction procedure

Before extraction of DL- α -tocopherol, quinoa seeds were ground using 1095 Knifetec Sample Mill.

Extraction procedure was carried out taking care to protect DL- α -tocopherol from light and oxidising conditions. Methanol was used as an extraction solvent. Approximately 5 g of sample was kept in 30 mL of extraction solvent in the dark at room temperature during the period of 16 h. Then it was sonicated in ultrasonic bath (VIMS elektrik, Loznica) for 40 min. After filtering through the wide-pore filter paper, methanol was evaporated using a Rota-vapor at 28 °C. Then 10 mL of n-hexan was added and sonicated again for 20 min. n-hexan was subsequently evaporated using a Rotavapor at 28 °C. The residue was dissolved in 1 mL of n-hexan and filtered (PTFE filter, 0.2 μ m pore size) in vial prior to HPLC analysis.

Reagents and standards

DL- α -tocopherol (analytical standard, min. 99.9%) for HPLC determination was obtained from Supelco Analytical, USA.

Methanol and n-hexan (HPLC grade) were purchased from Sigma-Aldrich.

Standard stock solution (concentration 1 mg/mL) was prepared by dissolving 10 mg of DL- α -tocopherol standard in 10 mL of isopropanol and was kept at a temperature of 4 °C protected from light.

The working standard solutions chosen for the calibration curve were prepared by dissolving the stock standard solution in n-hexan, in appropriate ratios, in order to obtain the following concentrations: 4, 10, 16, 22 and 28 μ g/mL. Dark volumetric flasks were kept protected from light at a temperature of 4 °C prior to HPLC analysis.

HPLC analysis

HPLC instrument was an Agilent 1200 system consisted of a UV-Visible detector (DAD), a binary pump, a vacuum degasser, an auto sampler. System control and data analysis were processed with Chemstation Software (Agilent Technologies). The chromatographic separation was performed on an Agilent column (Zorbax eclipse plus-C18, 1.8 μ m particle size, 100 mm x 2.1 mm I.D.) kept at 25 °C. The mobile phase consisted of 100% n-hexan, using an isocratic elution procedure with a flow-rate of 0.3 mL/min and a pressure of 105 bars. Five microliters of samples was injected onto the HPLC column. The total run time was 20 min. The chromatogram was monitored at a wavelength of a 285 nm during the experiment. The compounds were identified by chromatographic comparisons with authentic standards and against UV spectra comparison using a DAD detector. Quantification was based on the signal response, using the method of standard addition.

Method validation

The validation of the HPLC method for determination of DL- α -tocopherol in quinoa seeds was conducted by defining various validation parameters: linearity range, accuracy, precision and limits of detection (LOD) and quantification (LOQ).

Furthermore, robustness and specificity of the method were also investigated. The peaks were identified by their retention times, comparing the UV-Visible spectra and spiking with standards. Quantification has been done using an external standard

curve with five points. The linearity range was evaluated by plotting the peak area corresponding to analyte, as a function of the concentration introduced.

Precision of the method was determined under different circumstances, namely repeatability conditions, where independent experiments were carried out on the same day by the same operator using the same instrumentation (intra-day variation study), and intermediate conditions where the experiments were repeated on different days (inter-day variation study).

Accuracy was determined from the recovery rate at three concentration levels.

The limit of detection (LOD) and limit of quantification (LOQ) were relevant in the frame of this work, since the scope of the method was the determination of DL- α -tocopherol in quinoa seeds, due to relatively low content of this vitamin.

Data Analysis

The differences were considered to be significant when $p < 0.05$, and the data were analyzed using the software Statistica software version 12 (StatSoft inc., 2013).

RESULTS AND DISCUSSION

Linearity of calibration curve

Linearity of calibration curve was evaluated by the coefficient of determination (R^2) using linear regression analysis, testing series of six injections of five standard solutions of DL- α -tocopherol: 4, 10, 16, 22 and 28 $\mu\text{g/mL}$ (Fig. 1). The standard solutions were tested under above described chromatographic conditions. Calibration curve was constructed by linear regression of the peak area (Y) versus the concentration (X). Calibration curve showed good linearity ($R^2=0.9994$) of the DAD signal at 285 nm over a mentioned range of concentrations. The linear regression equation was as follows: $y = 5.6882x - 3.3233$.

Figure 1. Calibration curve of DL- α -tocopherol

Recovery and precision

Table 1.

Recovery and precision of the method

	Initial ($\mu\text{g/mL}$)	Added ($\mu\text{g/mL}$)	Found ($\mu\text{g/mL}$)	Recovery (%)	RSD (%)	
					Intra-day	Inter-day
Sample 1	37.47	4.00	4.2	105.0	0.5 ^a	1.2 ^a
Sample 2	37.47	8.00	7.7	96.25	0.7 ^a	1.5 ^a
Sample 3	37.47	12.0	12.8	106.7	0.8 ^a	0.9 ^a

Aim of the recovery study was to monitor the presence of matrix effects and to check the accuracy of the developed method. The accuracy of the analytical method was determined by spiking a known amount of DL- α -tocopherol standards into the samples of quinoa seeds and analyzing the percentage of recovery. Standard solutions were prepared in triplicate at three different levels (40%, 80% and 120%) of DL- α -tocopherol working standard (concentration 10 $\mu\text{g/mL}$). Each standard solution was injected six times. Recovery mean values for samples were within the 96.25% and 106.7% range, indicating good accuracy of the developed method (Table 1).

The precision of the method was expressed as the percentage of relative standard deviation (RSD%). In intra-day variation study ten injections of each of three standard solutions (concentrations 4.00 $\mu\text{g/mL}$, 8.00 $\mu\text{g/mL}$ and 12.0 $\mu\text{g/mL}$) were done three times per day (in the morning, afternoon and evening). In inter-day variation study, ten injections of the same three standard solutions were done once a day on three different days. The relative standard deviations ranged between 0.5% and 0.8% within intra-day variation study. Likewise the relative standard deviations were between 0.9% and 1.5% within inter-day variation study. Analysis of variance (ANOVA) showed that there were no significant differences ($p > 0.05$) between samples in intra-day and inter-day variation studies (Table 1).

The obtained results indicated that the proposed HPLC method has an acceptable precision of DL- α -tocopherol determination. Any interference was not present in the analyzed quinoa seeds.

Detection limit and quantification limit

The limits of detection (LOD) and quantification (LOQ) were defined as the lowest concentration of each compound, over five replicate injections with real sample extracts, which gave an average signal-to-noise ratio greater than 3 or 10, respectively (Cruz and Casal, 2013). The LOD and LOQ in this study were 3 ng/mL and 11 ng/mL of DL- α -tocopherol, respectively.

These values indicated that investigated HPLC method is sensitive to quantify DL- α -tocopherol in quinoa seeds.

Specificity

Specificity of the chromatographic method is the ability of the method to accurately measure the analyte response in the presence of all potential sample components (Shabir, 2003). The response of the analyte in quinoa seeds containing the analyte and all potential sample components (vitamins, lipids, minerals, proteins, etc.) is compared with the response of a solution containing only the analyte (Figure 2 and Figure 3).

Chromatogram of quinoa seeds' extract showed no peaks in the region in which DL- α -tocopherol eluted, indicating specificity of the method against interference. This observation is relevant for determination of DL- α -tocopherol, since it is present in food in micro amounts.

Robustness

The robustness of the method was examined by conducting the analysis under different chromatographic conditions, such as changes in HPLC flow rate or mobile phase solvent. Moreover, the time of ultrasonic processing after the extraction step was varied. Obtained results showed that the investigated varied parameters did not affect the retention time as well as the peak parameters. When comparing the peak of DL- α -tocopherol from standard solution with the peak of DL- α -tocopherol from extract of quinoa seeds, it can be noted that there was no modification in peak profile. This fact suggests that the method did not change with time or conditions.

CONCLUSIONS

This study focused on developing a method for accurate determination of the DL- α -tocopherol in quinoa seeds with adequate sensitivity, precision and accuracy, while being rapid and requiring only standard chromatographic equipment available to most analytical laboratories. The proposed method is suitable for DL- α -tocopherol quantification, enabling analysis of more samples in a short period of time. This method is thus suitable for routine a-

Figure 2. HPLC chromatogram corresponding to DL- α -tocopherol standard solution (concentration 28 $\mu\text{g/ml}$)

Figure 3. HPLC chromatogram corresponding to extract of quinoa seeds

analyses and can be used for quinoa seeds quality control or other related studies.

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РАЗВОЈ И ВАЛИДАЦИЈА МЕТОДЕ ЗА ОДРЕЂИВАЊЕ ДЛ- α -ТОКОФЕРОЛА У СЕМЕНУ КИНЕ (*Chenopodium quinoa* Willd.)

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Сажетак: У овом раду је развијена и валидирана HPLC метода за рутинско одређивање садржаја ДЛ- α -токоферола у семену киное. Поступак валидације заснива се на испитивању следећих параметара: линеарност, тачност, прецизност, граница детекције и граница квантификације, робустност и специфичност методе. Развијена метода се базира на екстракцији витамина ДЛ- α -токоферола применом ултразвучног купатила и метанола и хексана као екстракционих растварача. Нормално-фазна хроматографија под високим притиском са UV-VIS детектором коришћена је за детектовање испитиване супстанце. Метода је показала адекватну линеарност, док је релативна стандардна девијација износила између 0,5 и 0,8% (n=10). Осетљивост методе испитана је одређивањем границе детекције и границе квантификације које су износиле 3,0 ng/mL и 11,0 ng/mL. Такође, тестирањем робустности методе установљено је да метода није променљива при малим варијацијама параметара методе. Резултати су показали да је метода једноставна и тачна, и стога применљива у лабораторијама за одређивање садржаја ДЛ- α -токоферола у семену киное.

Кључне речи: ДЛ- α -токоферол, семе киное, HPLC

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